

Determination of Twenty-four β -Receptor Agonists in Chicken by Ultra-High-Performance Liquid Chromatography coupled with Triple Quadrupole Mass Spectrometry

No. LCMSMS-178E

Twenty-four β -receptor agonists were determined in chicken sample using Shimadzu Ultra-High-Performance Liquid Chromatograph Nexera X2 coupled with Triple Quadrupole Mass Spectrometer LCMS-8050. The calibration curves exhibited good linearity within the range of 0.1-10.0 $\mu\text{g/L}$ and the RSD of retention time and peak area was 0.07-0.40% and 0.44-4.12% respectively, indicating good instrument precision. The method detection limit and limit of quantitation was determined to be 0.004-0.025 $\mu\text{g/kg}$ and 0.014-0.082 $\mu\text{g/kg}$ respectively.

Keywords: Ultra-high-performance liquid chromatograph; Triple quadrupole mass spectrometer; Chicken; β -receptor agonists

■ Introduction

β -receptor agonists are a group of selective β_2 adrenergic receptor agonists. The compounds are named this way due to their ability to bind to most β -receptors on histiocyte membranes in animal body. They are a class of chemically synthesized phenylethanolamines that are structurally and physiologically similar to epinephrine and norepinephrine. The regulations in the EU 96/22/EC directive have prohibited the use of these drugs. Also, the Ministry of Agriculture and the State Administration of Quality Supervision of China have prohibited the use of such drugs in animal feed or on animals in the growing phase. Therefore, it is necessary to establish a fast and highly sensitive detection technique to quantify β -receptor agonists in chicken.

With reference to the method in "GBT 22286-2008: Determination of Multiple β -receptor Agonist Residues in Animal-Derived Food – LC - tandem MS Method", a UHPLC coupled with triple quadrupole MS method was developed for the determination of 24 β -receptor agonists in chicken. The use of Shimadzu Nexera X2 UHPLC and LCMS-8050 is used in this application note and the described method serves as a reference for relevant personnel.

■ Experimental

1.1. Instruments

Shimadzu Ultra-High-Performance Liquid Chromatograph (UHPLC) Nexera X2 coupled with Triple Quadrupole Mass Spectrometer LCMS-8050 was used. The specific configuration included LC-30AD \times 2 infusion pumps, DGU-20A_{SR} Online Degasser, SIL-30AC Autosampler, CTO-30AC Column Oven, CBM-20A System Controller, Triple Quadrupole Mass Spectrometer LCMS-8050, and LabSolutions Ver. 5.60 SP2 Chromatography Workstation.

1.2. Analytical Conditions

Liquid chromatography (LC) parameters

- Analyzer: Nexera X2
- Chromatographic column: Shimadzu Shim-pack XR-ODS III, 2.0 mm (I.D.) x 75 mm (L) x 1.6 μm
- Mobile phase: Phase A-0.1% formic acid solution; B-methanol
- Flow rate: 0.3 mL/min
- Injection volume: 5 μL
- Column temperature: 40°C
- Elution mode: gradient elution; the initial concentration of phase B was 5%, refer to Table 1 for gradient program.

Table 1: Gradient Program

Time(min)	Module	Command	Value
1.0	Pumps	Pump B Cone	5
4.0	Pumps	Pump B Cone	35
6.0	Pumps	Pump B Cone	90
8.0	Pumps	Pump B Cone	90
8.1	Pumps	Pump B Cone	5
10.0	Controller	Stop	

Mass spectrometry (MS) parameters

- Ion source: ESI, positive ion scan
- Nebulizing gas: 3 L/min
- Heating gas: 12 L/min
- Drying gas: 8 L/min
- Collision gas: Argon
- DL temperature: 250°C
- Heating module temperature: 400°C
- Scan mode: Multiple Reaction Monitoring (MRM)
- Dwell time: 15 msec
- Pause time: 3 msec
- MRM parameters: Refer to Table 2

Table 2: MRM Parameters

No.	Compound Name	CAS	Parent ion	Daughter ion	Q1 Pre Bias (V)	CE (V)	Q3 Pre Bias (V)
1	Cimaterol	54239-37-1	220.2	202.1* 143.1	-11 -26	-11 -23	-22 -27
2	Terbutaline	148-79-8	226.1	152.0* 125.1	-11 -26	-16 -23	-28 -24
3	Salbutamol	18559-94-9	240.0	222.1* 166.1	-12 -12	-11 -13	-24 -30
4	Cimbuterol	54239-39-3	234.2	143.1* 160.1	-27 -24	-25 -15	-26 -29
5	Fenoterol	13392-18-2	304.0	135.0* 286.0	-15 -15	-14 -18	-20 -25
6	Procaterol	72332-33-3	291.1	273.0* 231.2	-22 -15	-12 -21	-30 -26
7	Ritodrine	26652-09-5	288.0	270.1* 150.0	-14 -14	-13 -19	-30 -28
8	Clencyclohexerol	157877-79-7	319.1	203.0* 301.1	-15 -16	-14 -21	-21 -22
9	Hydroxymethyl-clenbuterol	38339-18-3	293.1	203.0* 275.1	-30 -15	-12 -19	-30 -22
10	Clenproperol	38339-11-6	263.1	245.0* 203.0	-13 -14	-12 -19	-27 -22
11	Clorprenaline	3811-25-4	214.1	154.1* 196.1	-21 -25	-13 -18	-21 -29
12	Clenbuterol	37148-27-9	277.1	203.0* 259.0	-28 -28	-11 -17	-28 -22
13	Formoterol	73573-87-2	345.0	149.0* 327.1	-17 -17	-14 -21	-23 -28
14	Bromochlorobuterol	78982-84-0	323.0	249.0* 305.0	-15 -15	-12 -18	-22 -27
15	Tulobuterol	41570-61-0	228.1	154.0* 118	-22 -26	-16 -28	-29 -22
16	Brombuterol	41937-02-4	367.0	349.0* 293.0	-18 -18	-13 -19	-25 -21
17	Mabuterol	56341-08-3	311.0	237.0* 293.0	-30 -15	-12 -18	-21 -26
18	Clenpenterol	37158-47-7	291.1	203.1* 273.1	-21 -22	-12 -16	-30 -22
19	Bambuterol	81732-65-2	368.0	294.1* 312.1	-18 -18	-15 -24	-22 -30
20	Clenisopenterol	157664-68-1	291.1	273.1* 188.1	-30 -15	-12 -22	-30 -21
21	Mapenterol	54238-51-6	325.1	237.0* 217.1	-16 -17	-17 -26	-26 -24
22	Phenylethanolamine A (PA)	1346746-81-3	345.1	150.1* 327.1	-18 -18	-24 -13	-28 -23
23	Clenhexerol	78982-88-4	305.1	287.1* 203.0	-30 -16	-14 -22	-20 -22
24	Salmeterol	89365-50-4	416.2	398.1* 380.1	-20 -20	-16 -20	-29 -27

Note: * represents quantifier ion

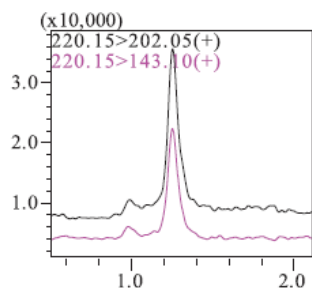
1.3. Sample Preparation

Sample was pretreated according to the sample pretreatment method stipulated in "GBT 22286-2008: Determination of Multiple β -receptor Agonist Residues in Animal-Derived Food – LC-Tandem MS Method."

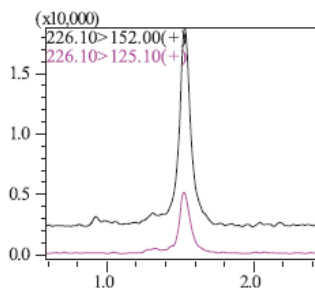
Preparation of standard solution: Mixed standard solutions containing 24 agonists at the concentration of 500 mg/L were prepared with methanol. This stock solution was sequentially diluted with methanol/water solution (5/95, V/V) to obtain a series of mixed standard working solutions at concentrations of 0.1, 0.5, 1.0, 5.0 and 10.0 $\mu\text{g/L}$.

■ Results and Discussion

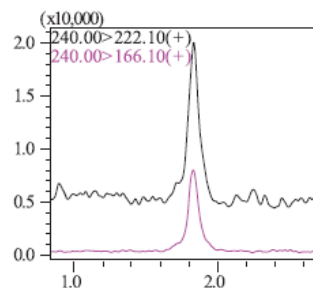
2.1. MRM Chromatograms of the Standard Samples



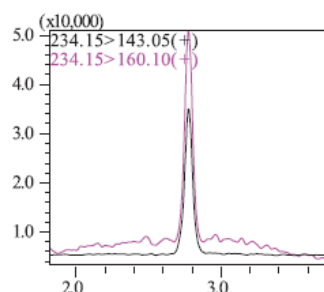
1. Cimaterol



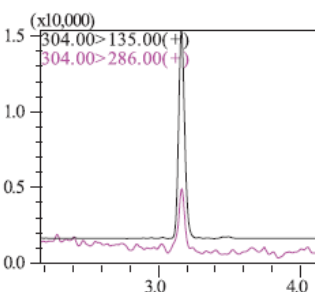
2. Terbutaline



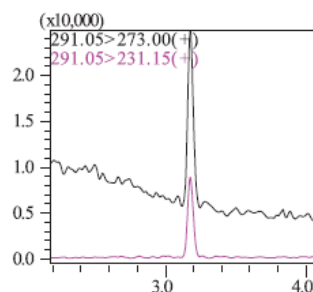
3. Salbutamol



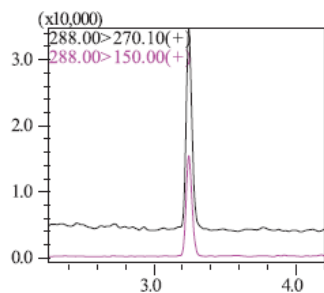
4. Cimbuterol



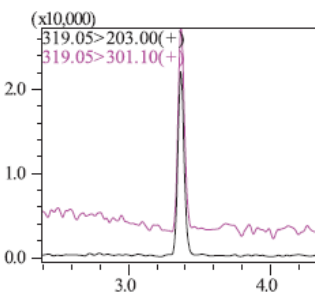
5. Fenoterol



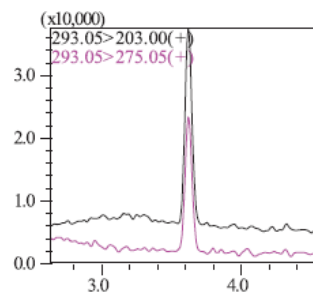
6. Procaterol



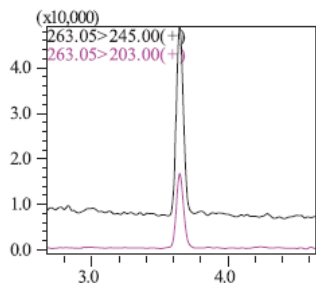
7. Ritodrine



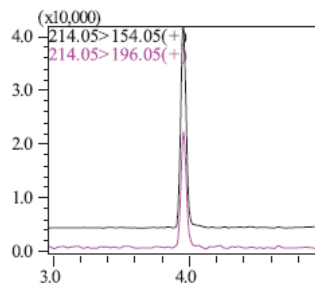
8. Clencyclohexerol



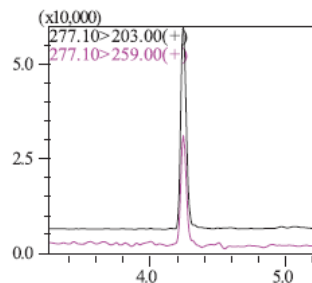
9. NA-1411



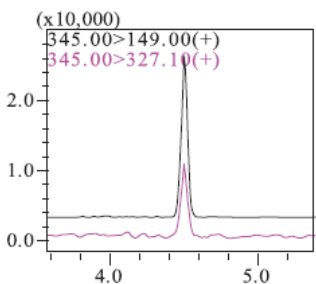
10. Clenproperol



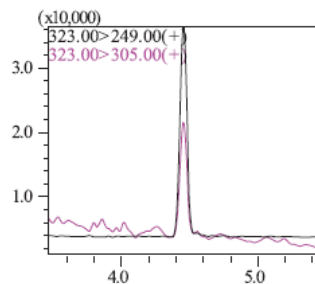
11. Clorprenaline



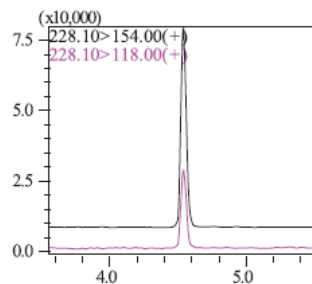
12. Clenbuterol



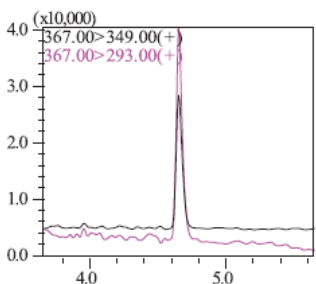
13. Formoterol



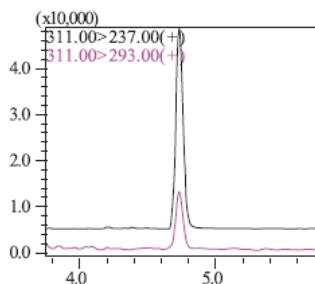
14. Bromchlorbuterol



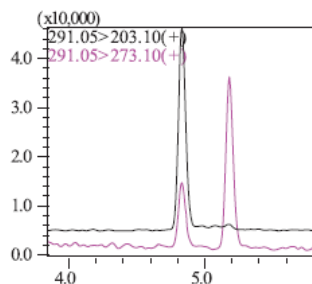
15. Tulobuterol



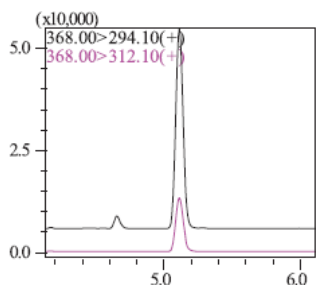
16. Brombuterol



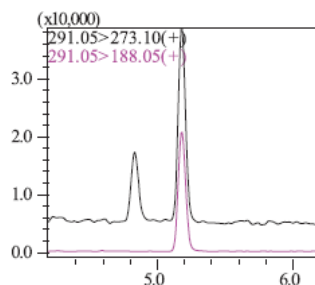
17. Mabuterol



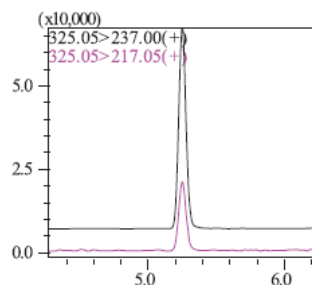
18. Clenpenterol



19. Bambuterol



20. Clenisopenterol



21. Mapenterol

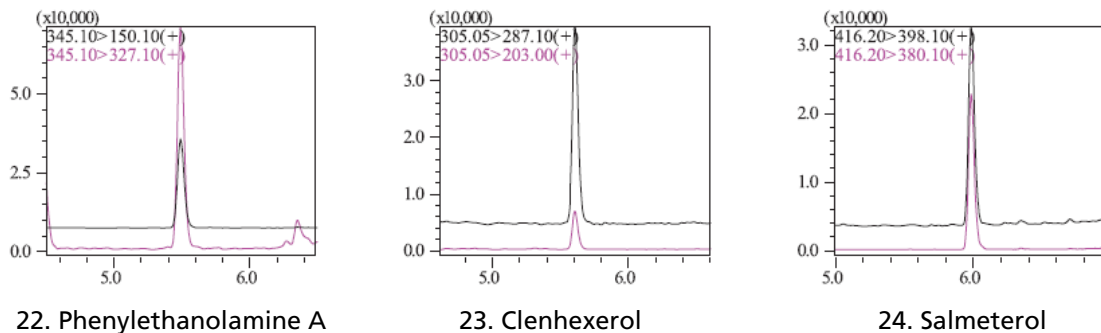


Figure 2: MRM Chromatograms of 24 Standard Samples at 0.5 µg/L

2.2. Calibration and Linearity

Calibration was performed using a 4-point curve with concentrations at 0.1, 0.5, 5 and 10 µg/L. External standard method was used to plot the calibration curves and the 24 compounds exhibited good linearity within the concentration range of 0.1-10 µg/L.

Some of the calibration curves are illustrated in Figure 3. The linear equations, correlation coefficients and linear range of all 24 compounds are shown in Table 3.

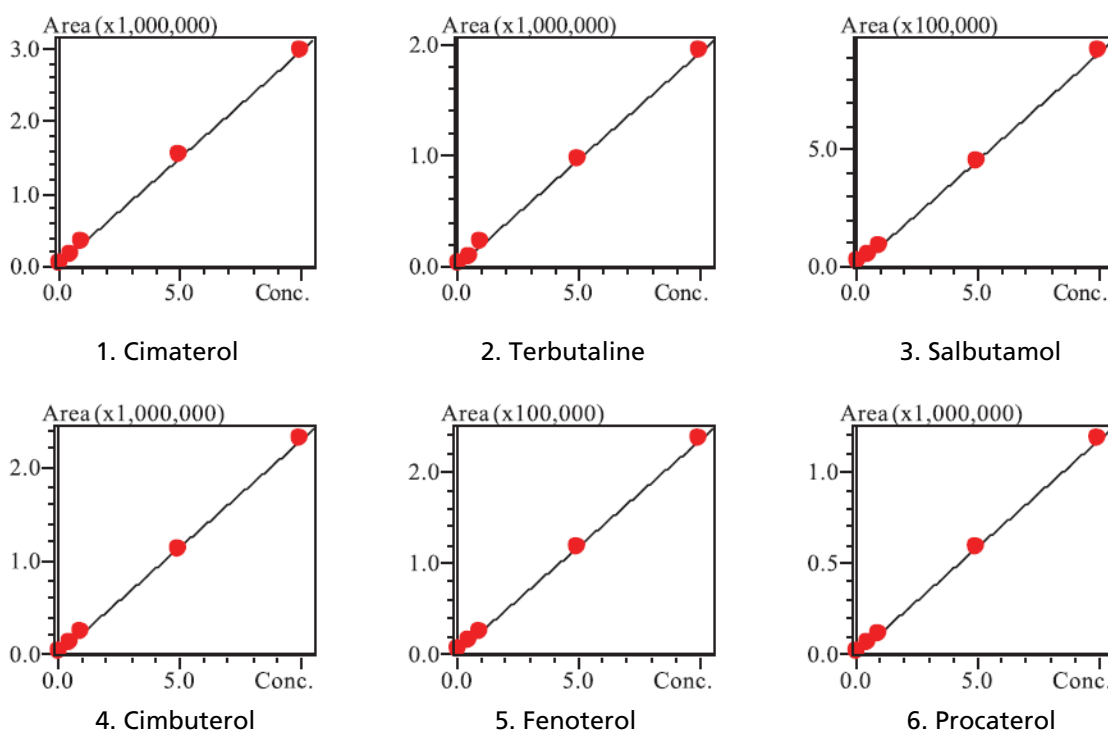


Figure 3: Calibration Curves of Cimaterol, Terbutaline, Salbutamol, Cimbuterol, Fenonterol and Procatamol

Table 3: Calibration Curve Parameters of 24 β -Receptor Agonists

No.	Compound Name	Calibration curve	Correlation coefficient (r)	Linear range ($\mu\text{g/L}$)
1	cimaterol	$Y = (295392)X + (22965.8)$	0.9996	0.1~10.0
2	terbutamol	$Y = (193158)X + (785.557)$	0.9997	0.1~10.0
3	salbutamol	$Y = (92096.7)X + (-4196.98)$	0.9990	0.1~10.0
4	cimbuterol	$Y = (228666)X + (49.6517)$	0.9993	0.1~10.0
5	fenoterol	$Y = (23336.1)X + (1754.60)$	0.9997	0.1~10.0
6	procaterol	$Y = (117874)X + (-1056.10)$	0.9997	0.1~10.0
7	ritodrine	$Y = (168469)X + (-2714.09)$	0.9998	0.1~10.0
8	clencyclohexerol	$Y = (140613)X + (6007.49)$	0.9981	0.1~10.0
9	NA	$Y = (142095)X + (9475.16)$	0.9997	0.1~10.0
10	clenproperol	$Y = (276702)X + (5397.33)$	0.9994	0.1~10.0
11	clorprenaline	$Y = (120122)X + (2530.85)$	0.9991	0.1~10.0
12	clenbuterol	$Y = (165991)X + (3818.39)$	0.9993	0.1~10.0
13	formoterol	$Y = (63694.2)X + (967.377)$	0.9991	0.1~10.0
14	bromchlorbuterol	$Y = (134267)X + (5363.64)$	0.9991	0.1~10.0
15	tulobuterol	$Y = (395663)X + (-10450.5)$	0.9997	0.1~10.0
16	brombuterol	$Y = (149063)X + (613.530)$	0.9995	0.1~10.0
17	mabuterol	$Y = (87975.6)X + (2167.41)$	0.9994	0.1~10.0
18	clenpenterol	$Y = (98120.2)X + (7456.52)$	0.9998	0.1~10.0
19	bambuterol	$Y = (110622)X + (-5098.32)$	0.9992	0.1~10.0
20	clenisopenterol	$Y = (241675)X + (3963.42)$	0.9996	0.1~10.0
21	mapenterol	$Y = (437827)X + (4879.97)$	0.9996	0.1~10.0
22	PA	$Y = (197828)X + (501.809)$	0.9996	0.1~10.0
23	clenhexerol	$Y = (213666)X + (5003.71)$	0.9996	0.1~10.0
24	salmeterol	$Y = (189691)X + (-5054.86)$	0.9999	0.1~10.0

2.3. Precision Experiment

The prepared mixed standard solutions at concentration of 0.1, 1.0 and 10.0 $\mu\text{g/L}$ were determined successively six times to examine instrument precision.

The repeatability results are shown in Table 4 and the RSD of retention time of standards at low-, mid- and high- concentrations was 0.07-0.40% and 0.44-4.12% respectively, indicating good instrument precision.

Table 4: Repeatability Results of Retention Time and Peak Area (n=6)

No.	Sample Name	RSD% (0.1 $\mu\text{g/L}$)		RSD% (0.1 $\mu\text{g/L}$)		RSD% (10.0 $\mu\text{g/L}$)	
		R.T	Area	R.T	Area	R.T	Area
1	cimaterol	0.40	3.90	0.24	2.18	0.29	1.08
2	terbutamol	0.35	2.93	0.13	2.44	0.23	1.17
3	salbutamol	0.11	3.85	0.19	1.87	0.18	2.49
4	cimbuterol	0.13	4.12	0.25	1.91	0.19	1.80
5	fenoterol	0.08	3.84	0.18	2.13	0.12	1.18
6	procaterol	0.07	2.74	0.19	1.95	0.08	0.44
7	ritodrine	0.13	3.80	0.12	2.88	0.09	0.87
8	clencyclohexerol	0.14	3.03	0.09	1.17	0.08	1.91
9	NA	0.11	2.10	0.12	2.09	0.13	2.13
10	clenproperol	0.16	2.87	0.10	1.80	0.14	1.85
11	clorprenaline	0.17	2.52	0.09	3.18	0.08	1.40
12	clenbuterol	0.15	3.27	0.14	1.44	0.10	1.76
13	formoterol	0.20	1.52	0.07	1.76	0.09	1.66
14	bromchlorbuterol	0.12	1.73	0.09	2.06	0.14	0.71
15	tulobuterol	0.10	2.78	0.07	1.71	0.07	1.47
16	brombuterol	0.09	2.12	0.10	1.47	0.08	1.11
17	mabuterol	0.14	2.83	0.18	1.61	0.09	2.03
18	clenpenterol	0.12	1.98	0.09	2.03	0.06	0.66
19	bambuterol	0.10	1.82	0.13	1.17	0.09	1.15
20	clenisopenterol	0.19	1.09	0.09	0.49	0.07	0.94
21	mapenterol	0.17	1.69	0.06	1.80	0.10	1.66
22	PA	0.14	0.69	0.09	1.18	0.14	1.57
23	clenhexerol	0.08	1.78	0.07	0.94	0.12	0.78
24	salmeterol	0.03	1.46	0.08	0.87	0.53	0.63

2.4. Method Performance

The matrix spike solution was prepared through dilution with chicken matrix to obtain a sample injection concentration of 0.1 µg/kg. The limit of detection (LOD, S/N=3) and limit of quantitation (LOQ, S/N=10) of these 24 β-Receptor Agonists substances are shown in Table 5.

Table 5: Limit of Detection and Limit of Quantitation of Twenty-four β-Receptor Agonists Substances

No.	Compound name	LOD (µg/kg)	LOQ (µg/kg)
1	cimaterol	0.010	0.034
2	terbutamol	0.006	0.020
3	salbutamol	0.025	0.082
4	cimbuterol	0.020	0.067
5	fenoterol	0.009	0.030
6	procatamol	0.017	0.056
7	ritodrine	0.011	0.037
8	clencyclohexerol	0.021	0.069
9	NA	0.015	0.051
10	clenproperol	0.010	0.032
11	clorprenaline	0.012	0.039
12	clenbuterol	0.009	0.031
13	formoterol	0.012	0.040
14	bromchlorbuterol	0.004	0.014
15	tulobuterol	0.006	0.020
16	brombuterol	0.011	0.036
17	mabuterol	0.008	0.027
18	clenpenterol	0.012	0.040
19	bambuterol	0.009	0.030
20	clenisopenterol	0.010	0.033
21	mapenterol	0.006	0.019
22	PA	0.004	0.015
23	clenhexerol	0.012	0.038
24	salmeterol	0.010	0.034

2.5. Matrix Spike and Recovery Experiment

After sample pretreatment as stated in 1.3, mixed standard stock solution was added to the blank matrix to obtain a final concentration of 1.0 µg/kg. The recovery and RSD of the 3 replicates were determined. The results are shown in Table 6, and the matrix spike recovery was in the range of 90.1-107%. The chromatogram of the blank matrix, and matrix spike sample are shown in Figures 4 and 5 respectively.

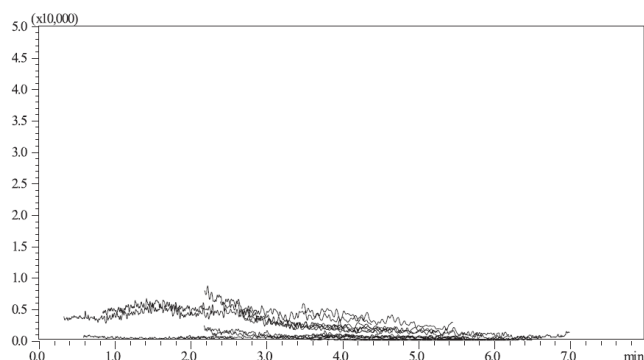


Figure 4: Chromatogram of the Blank Matrix

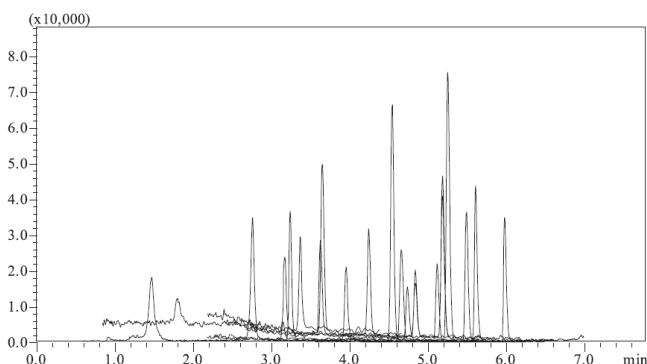


Figure 5: Chromatogram of the Matrix Spike Sample

Table 6: Recovery Results of the Matrix Spike Sample (n=3)

No.	Compound name	Recovery (%)	RSD%	No.	Compound Name	Recovery (%)	RSD%
1	Cimaterol	102.4	2.01	13	formoterol	95.3	1.93
2	terbutamol	104.7	2.09	14	bromchlorbuterol	93.1	1.71
3	salbutamol	94.8	2.02	15	tulobuterol	96.3	1.85
4	cimbuterol	91.5	1.02	16	brombuterol	94.8	0.95
5	fenoterol	99.5	2.04	17	mabuterol	102.5	1.63
6	procatamol	96.8	0.65	18	clenpenterol	99.5	1.17
7	ritodrine	98.5	1.11	19	bambuterol	96.1	1.58
8	clencyclohexerol	90.1	0.16	20	clenisopenterol	98.5	0.91
9	NA	98.6	0.74	21	mapenterol	99.1	1.53
10	clenproperol	101.9	0.92	22	PA	103.9	0.53
11	clorprenaline	97.2	0.63	23	clenhexerol	95.4	1.06
12	clenbuterol	94.1	1.14	24	salmeterol	100.3	1.79

■ **Conclusion**

The described method demonstrates the determination of twenty-four β -receptor agonists in chicken using Shimadzu Nexera X2 UHPLC coupled with Triple Quadrupole Mass Spectrometer LCMS-8050. This method successfully separates and quantifies 24 β -receptor agonists in 10 minutes. The RSD of retention time and peak area was 0.07-0.40% and 0.44-4.12% respectively, indicating good instrument precision. For all 24 β -receptor agonists, the calibration curves show good linearity with correlation coefficient greater than 0.9981. The LOD and LOQ was 0.004-0.025 $\mu\text{g}/\text{kg}$ and 0.014-0.082 $\mu\text{g}/\text{kg}$ respectively. With characteristics such as rapid, good repeatability and high sensitivity, this method demonstrates to be suitable for the detection of β -receptor agonists in chicken.

UAFMS

ULTRA FAST MASS SPECTROMETRY



LCMS-8040



LCMS-8045



LCMS-8050



LCMS-8060



LCMS-2020



Q-TOF LCMS-9030

Founded in 1875, Shimadzu Corporation, a leader in the development of advanced technologies, has a distinguished history of innovation built on the foundation of contributing to society through science and technology. Established in 1975, Shimadzu Scientific Instruments (SSI), the American subsidiary of Shimadzu Corporation, provides a comprehensive range of analytical solutions to laboratories throughout North, Central, and parts of South America. SSI maintains a network of nine regional offices strategically located across the United States, with experienced technical specialists, service and sales engineers situated throughout the country, as well as applications laboratories on both coasts.

For information about Shimadzu Scientific Instruments and to contact your local office, please visit our Web site at www.ssi.shimadzu.com



Shimadzu Corporation
www.shimadzu.com/an/

SHIMADZU SCIENTIFIC INSTRUMENTS, INC.
Applications Laboratory
7102 Riverwood Drive, Columbia, MD 21045
Phone: 800-477-1227 Fax: 410-381-1222
URL <http://www.ssi.shimadzu.com>

For Research Use Only. Not for use in diagnostic procedures. The content of this publication shall not be reproduced, altered or sold for any commercial purpose without the written approval of Shimadzu. The information contained herein is provided to you "as is" without warranty of any kind including without limitation warranties as to its accuracy or completeness. Shimadzu does not assume any responsibility or liability for any damage, whether direct or indirect, relating to the use of this publication. This publications is based upon the information available to Shimadzu on or before the date of publication, and subject to change without notice.

© Shimadzu Scientific Instruments, 2018
First Edition: August 2018