

# Metabolite profiling applied to biomarker discovery in pancreatic cancer using high resolution LC-MS/MS

Alan Barnes<sup>1</sup>; Emily G Armitage<sup>1</sup>; Neil Loftus<sup>1</sup>; Elon Correa<sup>2</sup>; Lynne Howells<sup>3</sup>; S n Takeda<sup>4</sup>; Wen Chung<sup>5</sup>

<sup>1</sup>Shimadzu Corporation, Manchester, UK; <sup>2</sup>Liverpool John Moores University, Liverpool, UK; <sup>3</sup>Institute for Precision Health, The University of Leicester, UK; <sup>4</sup>Department of Anatomy, Teikyo University School of Medicine, Tokyo, Japan; <sup>5</sup>Leicester HPB Unit, Glenfield Hospital, Leicester, UK

## Overview

- High resolution LC-MS/MS QTOF analysis was applied to pancreatic adenocarcinoma (PDAC) samples, a disease characterised by early metastasis and low survival rate.
- Previously published and new biomarkers were identified in this work; notably elevated bile acids and reduced LPEs, LPCs and PCs.
- Comparison to direct probe electrospray ionisation data provided added confidence in biomarker identification that may be considered in the future as part of a screening system for early detection of PDAC.

## 1. Introduction

In the present study, two metabolite profiling methodologies were used to detect metabolite changes between patient serum samples with pancreatic ductal adenocarcinoma (PDAC) and healthy controls; a standardised approach in LC-MS/MS based metabolite profiling and a direct analysis method (Direct Probe Ionisation Mass Spectrometry – DPiMS<sup>1</sup>). In this study high resolution LC-MS/MS was used to characterise the metabolic profiles of serum from patients with PDAC compared to healthy controls to reveal key metabolic differences in these phenotypes and confirm DPiMS measurements.

## 2. Materials and Methods

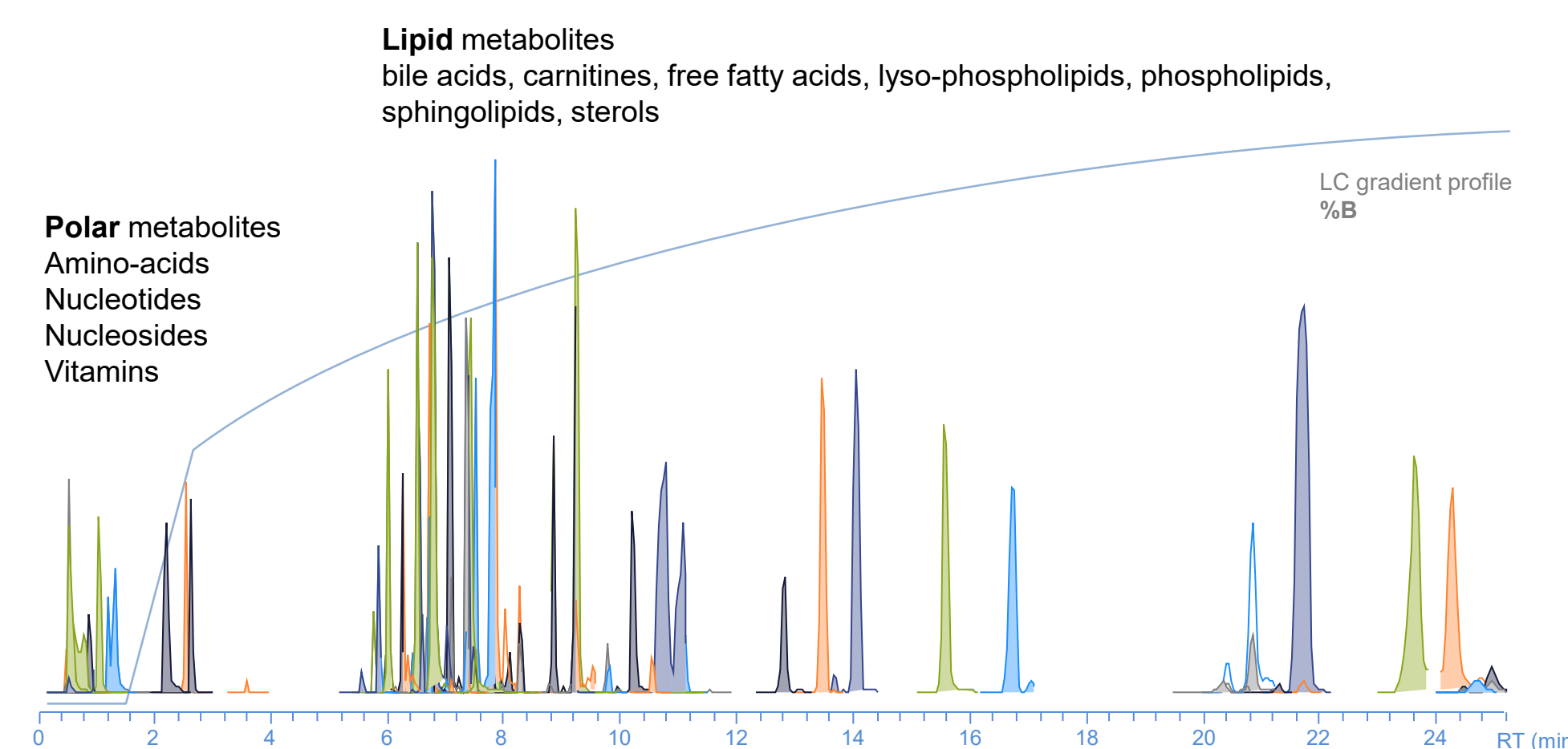
Serum samples included healthy controls (n=30) and PDAC (n=30). The research project was approved by the Pancreatic Cancer Research Fund Tissue Bank (PCRFTB) Tissue Access Committee.

- Reverse phase LC Separation.**
  - Acquity C18 BEH (2.1x100mm 1.7 m); 50 C, flow rate 0.4 mL/min
  - Binary gradient; water + 0.1% formic acid, and acetonitrile + 0.1% formic acid
  - Cycle time 35 minutes.
- LC-MS/MS Mass Spectrometry Detection.** High resolution QTOF LC-MS/MS analysis (LCMS-9030, Shimadzu Corporation, Japan) using external mass calibration for positive and negative mode ESI. The untargeted metabolite profiling method used a TOF MS mass scan followed by a series of DIA-MS/MS mass scans.
  - TOF MS mass scan m/z 60-1000; 100 msec
  - DIA-MS/MS mass scans m/z 40-1000; 33 msec for each precursor isolation window; isolation width 35 Da; collision energy spread 5-55V; 27 mass scan events. Scan cycle time 0.99 second (28 mass scans in total).
- DPiMS Mass Spectrometry Detection.** High resolution QTOF MS/MS analysis (DPiMS QT, Shimadzu Corporation, Japan); direct sample analysis (no LC).
  - TOF MS mass scan m/z 100-1500; 100 msec
  - For DPiMS compound identification, DIA-MS/MS mass scans m/z 100-1500; collision energy spread 5-55V, precursor isolation window 1 Da, positive and negative

- Data processing.** Several data processing tools are available on-line for feature detection, alignment and identification such as MS-DIAL, MZmine3, GNPS, XCMSonline plus. This study used MS-DIAL for peak processing and alignment, MetaboAnalyst for statistical analysis and LabSolutions Insight for metabolite identification.

## 2.1 Applying a generic reverse phase LC-MS/MS method

The method has been optimised for a broad range of metabolite classes

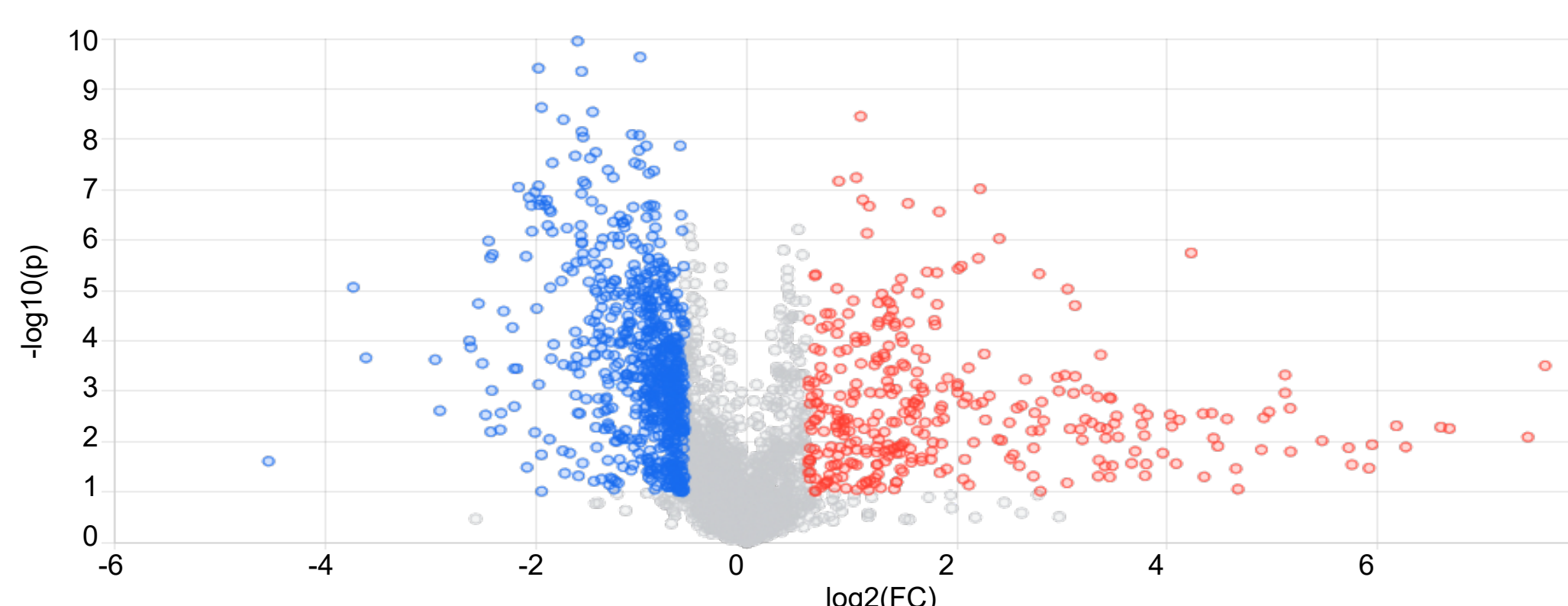


**Figure 1.** Separation of a broad range of metabolites using a single run reverse phase method with MS and DIA-MS/MS detection. In this study, the primary focus of the investigation was to profile the change in lipid distributions in PDAC samples compared to controls.

## 2.2 Data Processing Workflow for LC-MS/MS

One approach for processing untargeted metabolomics data;

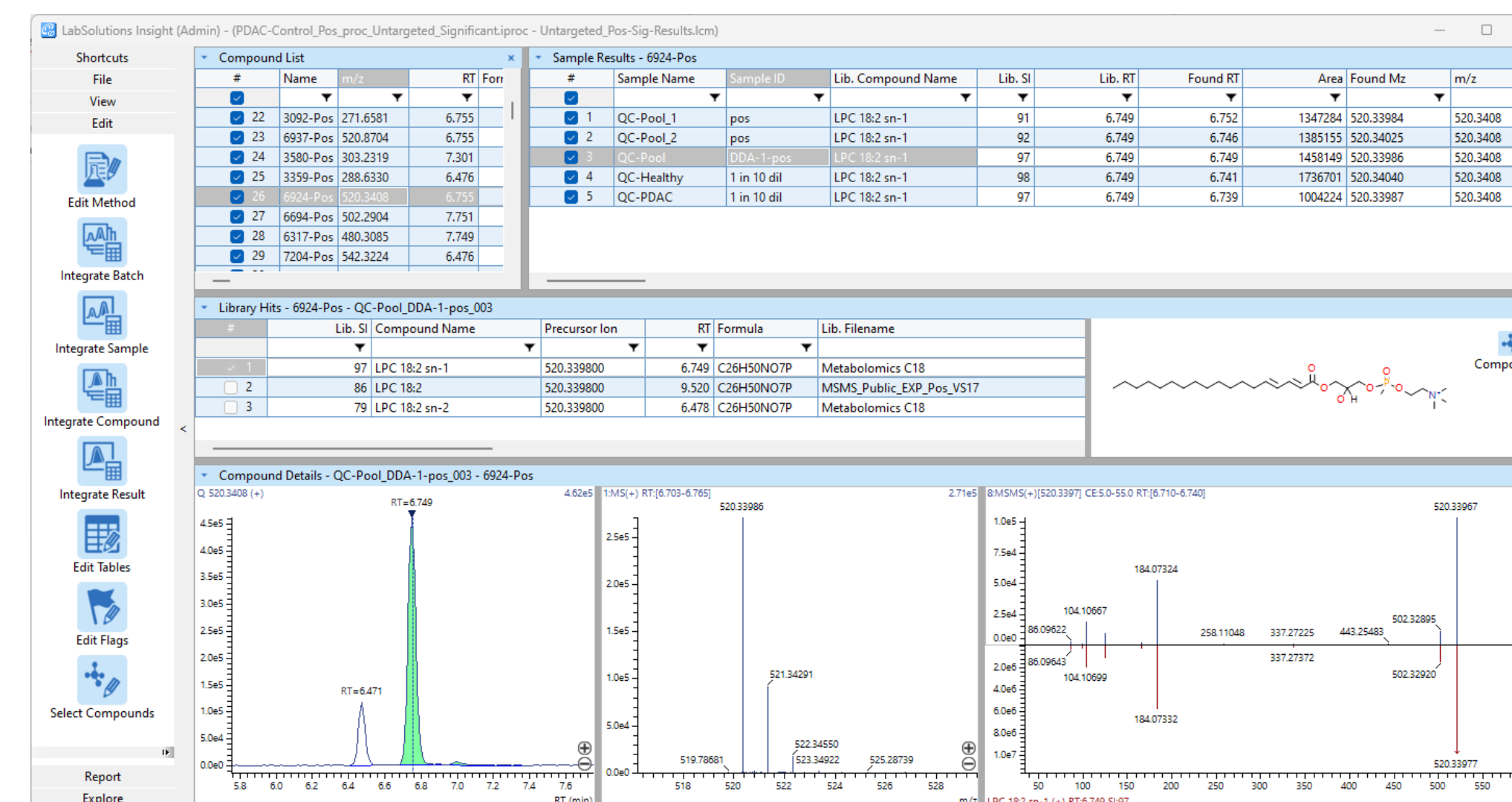
- MS-DIAL** | raw data file processing, feature detection, alignment, filters applied.
- MetaboAnalyst 5.0** | univariate statistical analysis, volcano plot.
- Compound Identification** | LabSolutions Insight.



**Figure 2.** Volcano plot showing features extracted using HRMS LC-MS in positive and negative ion mode that were significantly increased or decreased in PDAC serum samples compared to healthy controls (fold change >1.5, p<0.1).

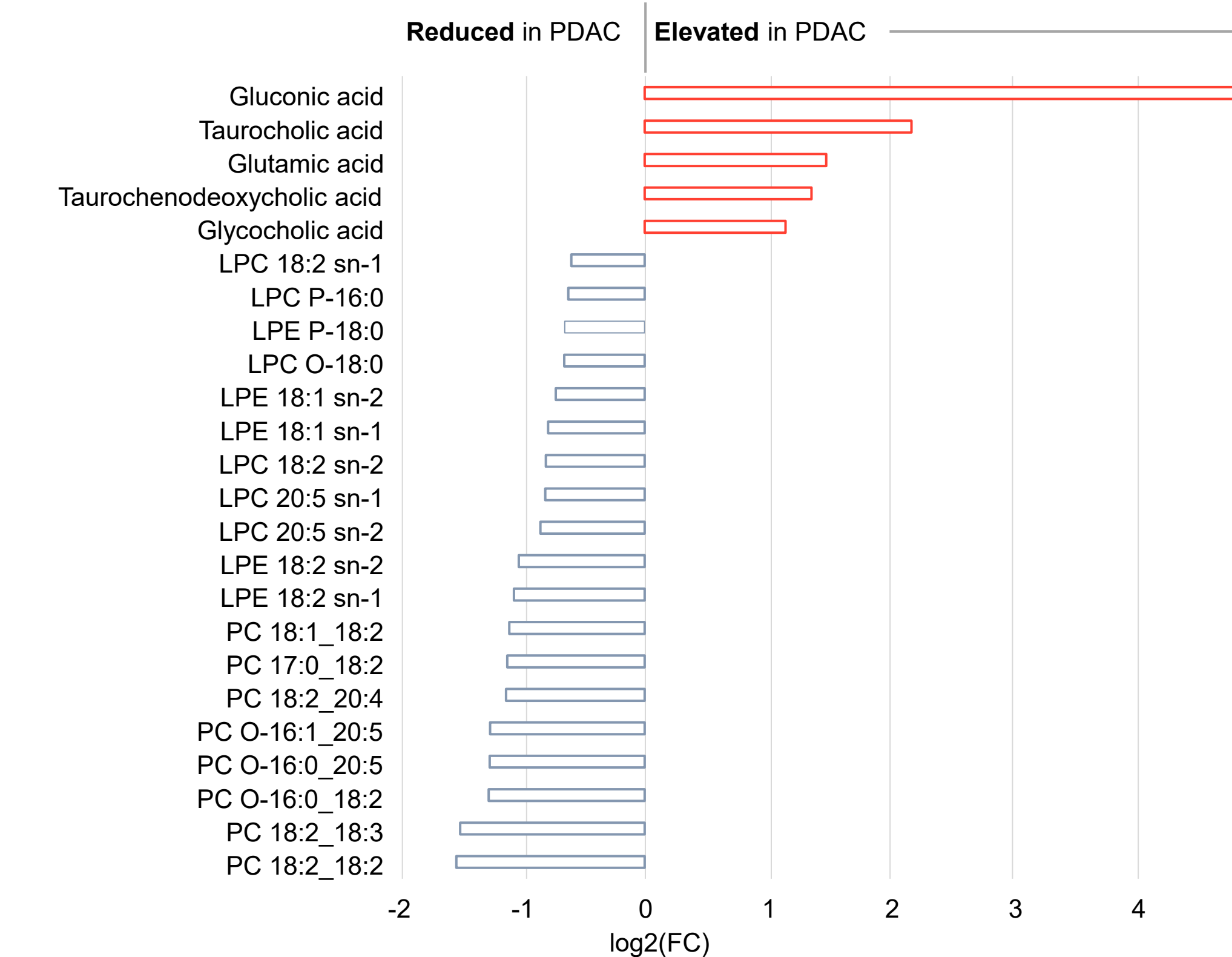
MS-DIAL generated an aligned data array that was subsequently filtered (metabolite feature was present in >50% of the QCs with QC RSD<20% and present in at least 80% of either the healthy or PDAC group).

## 2.3 Metabolite Identification using LC-MS/MS



**Figure 3.** LabSolutions Insight software application was used for metabolite identification. DIA and DDA-MS/MS mass spectra were searched against an in-house development metabolomics library and the MS-DIAL MSP spectral kit (<http://prime.psc.riken.jp/compms/msdial/main.html>).

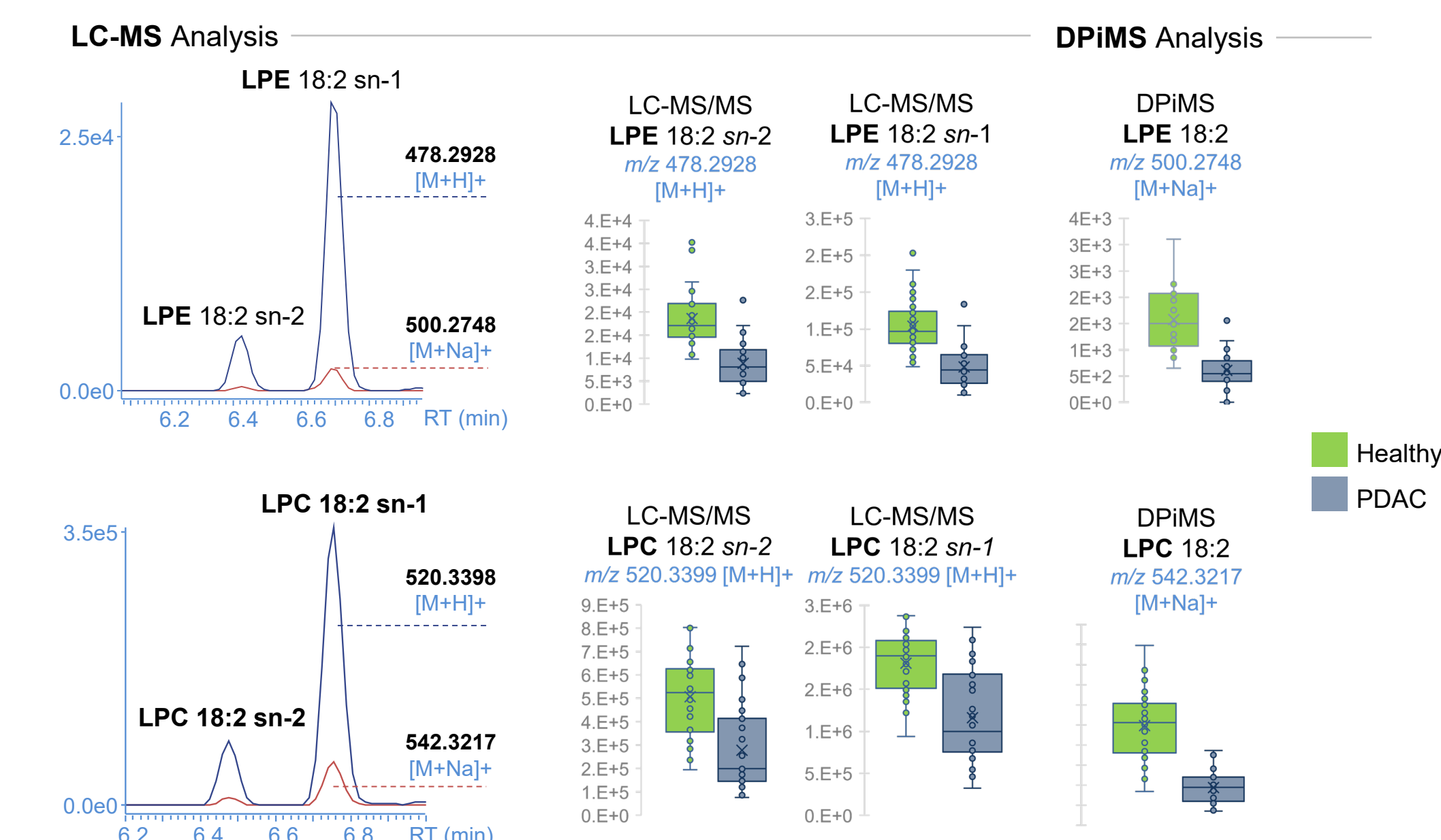
## 3. Results



**Figure 4.** Bar chart highlighting the most significant changes (fold change >1.5) in PDAC samples relative to healthy controls that could be annotated at the MSMS level (Metabolomics Standards Initiative level 1 or 2).

## 3.1 HR LC-MS/MS comparison to PESI

Relative levels of biomarker candidates were compared between different acquisition methods; liquid chromatography electrospray ionisation (LC-ESI) and direct probe ionisation mass spectrometry (also known as probe electrospray ionisation (PESI)). Both high-resolution LC-MS/MS data and DPiMS showed the same direction of change.



**Figure 5.** LC-MS/MS and DPiMS results for two lipids; LPE 18:2 and LPC 18:2. DPiMS shows the same direction of change as LC-MS/MS for both lipids, however, DPiMS cannot resolve the sn-1 and sn-2 isomers. As DPiMS is a direct analysis technique, adducts are often the dominant ion species (adducts are also present in LC-MS/MS, but the molecular ion is typically the dominant ion).

## 4. Conclusions

- High resolution LC-MS/MS detected a panel of potential biomarkers that may be used to differentiate between PDAC serum samples and healthy controls.
- Metabolites were identified to MSI level 1 or 2 using an in-house metabolomics library and the MS-DIAL MSP spectral kit (<http://prime.psc.riken.jp/compms/msdial/main.html>).
- Analysis by LC-MS/MS helped confirm candidate biomarker identification from DPiMS providing verification to a previous nominal mass study<sup>2</sup>.

## 5. References

- Mandel et al. Application of Probe Electrospray Ionization Mass Spectrometry (PESI-MS) to Clinical Diagnosis: Solvent Effect on Lipid Analysis. J. Am. Soc. Mass Spectrom. 2012, 23, 11, 2043–2047
- Chung et al. Using probe electrospray ionization mass spectrometry and machine learning for detecting pancreatic cancer with high performance. Am J Transl Res. 2020; 12(1): 171–179.