

Simplifying dioxins analysis in foods and feeds using GC-MS/MS in compliant with the EU Commission Regulation

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Overview

- ◆ To enable analysis of dioxins in food and animal feed products, a versatile and sensitive GC-MS/MS equipment is required.
- Shimadzu GCMS-TQ8050 NX system (Fig. 1) equipped with AOC-30i + 20sU and Boost Efficiency Ion Source (BEIS) demonstrated to achieve low femtogram levels by SPL injector at 1µL for the analysis of dioxins in accordance with EU Commission Regulation 589/204 and 644/2017.
- Eurofins Special Tests, one of the most important laboratories for dioxins analysis in Brazil, collaborated with this work.



Fig. 1 Shimadzu GCMS-TQ8050 NX.

1. Introduction

- Dioxins in food and animal feed products are one of the most important Persistent Organic Pollutants (POPs) to evaluate. Quantitative analysis of dioxins in low concentration levels is necessary due to their high toxicity. Regarding to the analysis, EU Commission Regulations stablish analysis methods using GC-MS/MS as official test methods with the same positioning as methods using GC-HRMS.
- In addition to the importance to monitor them, analysis of dioxins by GC-MS/MS has attracted interest. Due to the complexity of matrix and low limits, it is common to use a PTV injector by Large Volume Injection technique to reach femtogram levels. Therefore, a robust and sensitive method that uses minimal injection volume is desirable.

2. Methods

• For the various food samples, pretreatment was performed using Eurofins Special Tests internal procedure.

• For the analytical parameters, the conditions registered in the Shimadzu's method package named EU Regulation Compliant GC-MS/MS Method Package for Dioxins in Foods (Fig. 2) were used. Some changes were performed to be in accordance with Eurofins internal requirement.

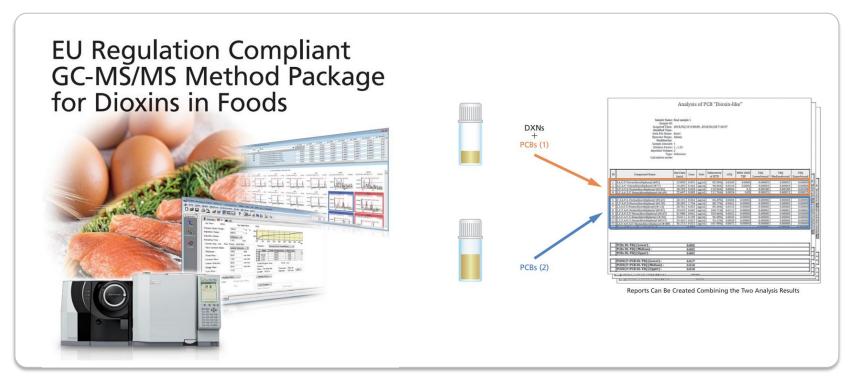


Fig. 2 Shimadzu's EU Regulation Compliant GC-MS/MS method package for dioxins in foods.

• The retention times for all dioxins and furans congeners were done automatically using the Automatic Adjustment of Retention Time (AART) function.

◆ Also, the use of hydrogen as alternative gas was evaluated in direct comparison with results already obtained using another instrument.

3. Result

All calibration curve data for each congeners were obtained from $25-10000 \text{fg/}\mu\text{L}$, as shown in **Table 1**.

• Considering the maximum permitted concentrations (ML), the ML for pig's fat and meat were the lowest at 1pg/g of fat (sum of dioxins, WHO-PCDD/F-TEF). The limit of quantification (LOQ) was considered 1/5 of ML

• The precision and accuracy parameters were conducted spiking blank samples at LOQ and ML levels. For all injections, the recovery results were obtained within range of 60-120%. The %RSD at 0.2pg/g was 6.6 and at 1.0pg/g was 8.2. The validation procedures were conducted considering replicates (n=20).

| Iable 1 Calibration curve data for each congeners | | | | | | | | |
|---|---------|---------|----------|----------|-----------|------------|----------------|--|
| Compound | 25fg/µL | 50fg/µL | 100fg/µL | 500fg/µL | 2000fg/µL | 10000fg/µL | R ² | |
| 2,3,7,8-TCDD | 23.726 | 47.068 | 98.992 | 611.919 | 2132.035 | 9968.025 | 0.99972 | |
| 1,2,3,7,8-PeCDD | 31.398 | 49.623 | 109.694 | 546.798 | 2079.636 | 9981.622 | 0.99993 | |
| 1,2,3,4,7,8-HxCDD | 32.870 | 54.467 | 107.803 | 511.173 | 2202.076 | 9958.906 | 0.99955 | |
| 1,2,3,6,7,8-HxCDD | 28.251 | 69.423 | 98.808 | 527.952 | 2136.168 | 9971.275 | 0.99980 | |
| 1,2,3,7,8,9-HxCDD | 26.323 | 63.742 | 137.118 | 593.647 | 2218.754 | 9951.124 | 0.99950 | |
| 1,2,3,4,6,7,8-HpCDD | 22.205 | 53.492 | 99.745 | 526.244 | 2059.510 | 9986.778 | 0.99996 | |
| OCDD | 47.728 | 106.675 | 231.714 | 1197.367 | 4429.895 | 19903.808 | 0.99947 | |
| 2,3,7,8-TCDF | 33.147 | 66.368 | 109.577 | 647.276 | 2220.264 | 9948.385 | 0.99940 | |
| 1,2,3,7,8-PeCDF | 38.081 | 51.477 | 91.500 | 529.228 | 2144.940 | 9969.596 | 0.99977 | |
| 2,3,4,7,8-PeCDF | 36.565 | 60.270 | 106.934 | 608.053 | 2161.619 | 9962.124 | 0.99968 | |
| 1,2,3,4,7,8-HxCDF | 33.063 | 70.965 | 112.130 | 569.740 | 2198.799 | 9956.507 | 0.99959 | |
| 1,2,3,6,7,8-HxCDF | 36.116 | 62.819 | 126.985 | 575.487 | 2233.598 | 9949.144 | 0.99944 | |
| 2,3,4,6,7,8-HxCDF | 34.946 | 59.856 | 101.504 | 526.507 | 2184.400 | 9961.705 | 0.99964 | |
| 1,2,3,7,8,9-HxCDF | 35.712 | 51.610 | 109.066 | 589.710 | 2191.344 | 9957.120 | 0.99959 | |
| 1,2,3,4,6,7,8-HpCDF | 36.548 | 64.110 | 106.874 | 525.396 | 2183.628 | 9961.836 | 0.99965 | |
| 1,2,3,4,7,8,9-HpCDF | 38.228 | 47.944 | 109.585 | 533.886 | 2291.373 | 9939.912 | 0.99906 | |
| OCDF | 59.825 | 122.586 | 208.172 | 1142.006 | 4566.173 | 19879.446 | 0.99912 | |

◆ To evaluate accuracy by proficiency tests, one assay was conducted in pork liver achieving a Z-score at 0.4.

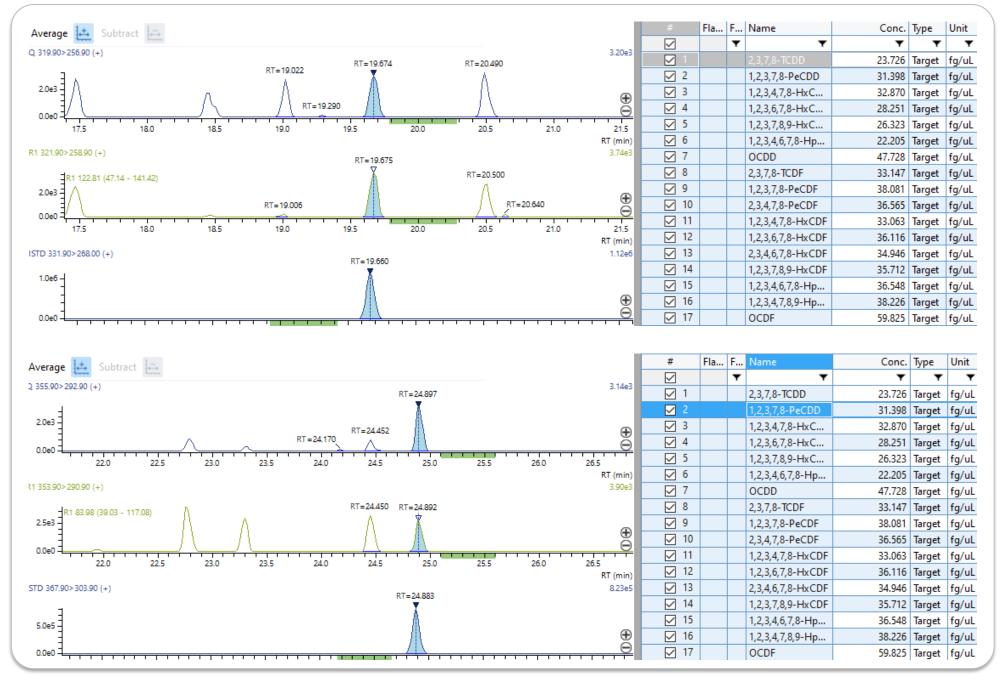


Fig. 3 Chromatograms for 2,3,7,8-TCDD and 1,2,3,7,8-PeCDD at 25fg/µL

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Table 1 Calibration curve data for each congeners

• Fig. 3 and Fig. 4 represent chromatograms for some congeners.

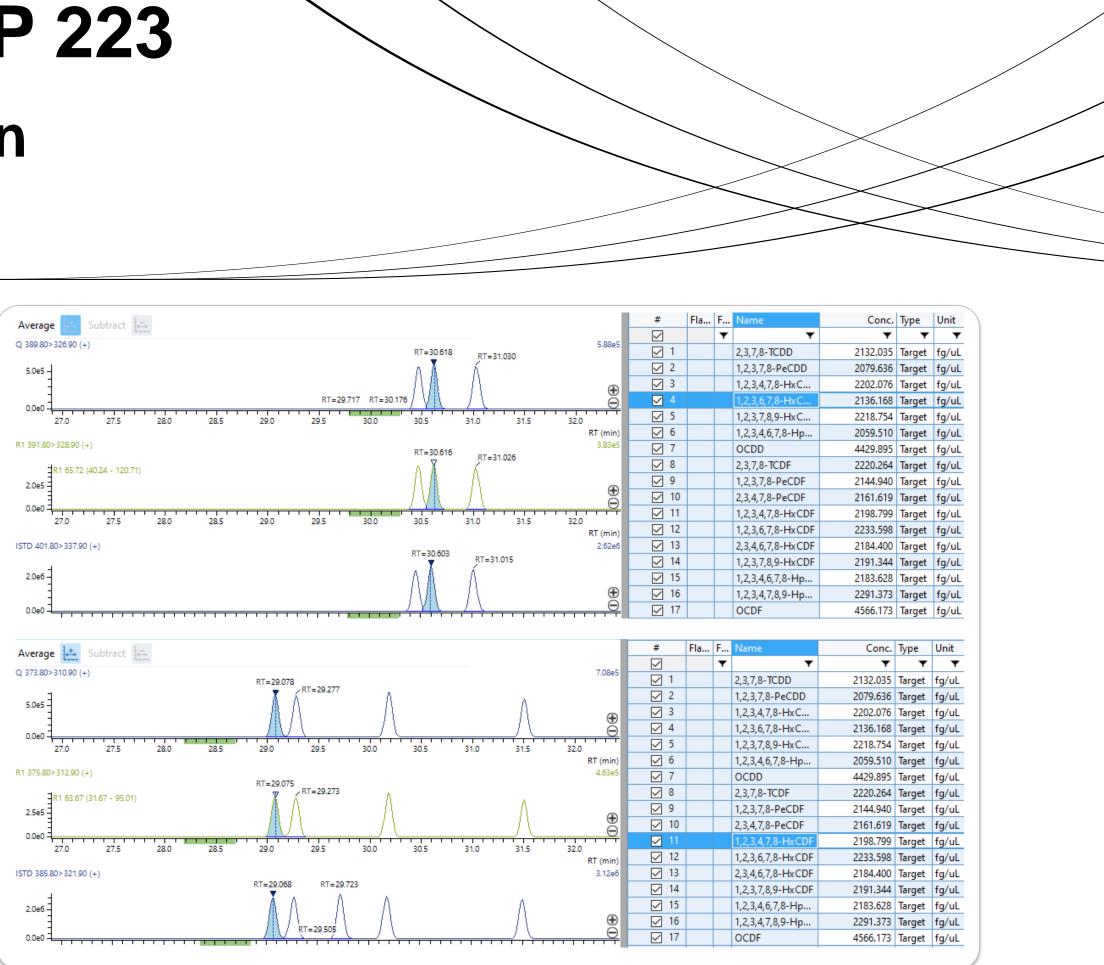


Fig. 4 Chromatograms for 1,2,3,6,7,8-HxCDD and 1,2,3,4,7,8-HxCDF at 2000fg/µL

groups by He and H_2 .

Table 2 Sensitivity assessment for LOQ comparing data obtained by He
 and H_2 .



4. Conclusion

- durability of the instrument.
- analysis in 23.4%.

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◆ Table 2 represents summary of sensitivity (LOQ) for congeners

| Concentration (fg/µL) | | | |
|-----------------------|--|--|--|
| LOQ (He) | LOQ (H ₂) | | |
| 25 | 25 | | |
| 25 | 25 | | |
| 25 | 250 | | |
| 25 | 500 | | |
| 25 | 500 | | |
| | LOQ (He) 25 25 25 25 25 | | |

 \checkmark Regarding the robustness of instrument using low injection volume, more than 6900 injections were done maintaining sensitivity, showing

✓ The groups TCDD/F and PeCDD/F were the aim of this study, since they have the highest TEF (TEF=1). Until penta congeners, it was possible to achieve the same LOQ by helium as carrier gas. The other congeners had higher LOQ. An important result was the reduction of