

MS Imaging Spectrum Annotation Tool using Accurate Mass and Isotope Pattern Analysis

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Introduction

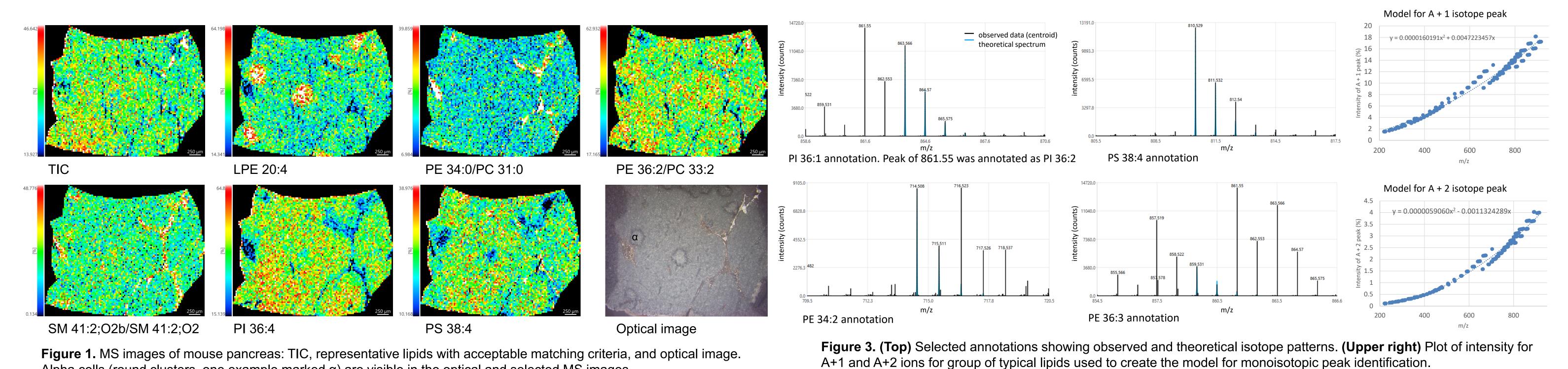
MS Imaging reveals the spatial distribution of important biomolecules to discover the details of disease processes, find drug distribution patterns, or characterize new materials. Real-world samples can result in complex mass spectra which are time-consuming to process manually. Searching for targets may turn up false matches, even having less than one part-per-million mass error. Using isotope pattern can exclude false matches and speed up processing. We developed a new MS spectrum annotation tool using accurate mass and isotope pattern analysis to quickly match targets to data.

Methods

Tissue samples were prepared by vapor-depositing 1,5-Diamino-naphthalene onto mouse pancreas affixed to conductive glass slides using a Shimadzu iMLayerTM sublimation unit. Then tissues were analyzed by MALDI MS imaging using a Shimadzu IMScope QTTM, composed of an ionization unit connected to a QTof mass spectrometer. Spectra were exported from the acquisition software and analyzed by a newly developed data processing tool.

Setting	Value
Pitch X	30.0 μm
Pitch Y	30.0 μm
Polarity	Negative
Scan Range (m/z)	500 to 1000
Number of Pixels	8752
Laser Irradiation Number	80 shots
Laser Repetition Frequency	2000 Hz
Laser Diameter	Approximately 35 μm
Laser Intensity	65%
DL Temperature	250 deg C

Table 1. Imaging MS settings optimized for mouse pancreas tissue, DAN coated, in negative MS mode.



MS resolving power:

Mass error threshold (ppm):

Intensity threshold (counts):

A-Number and Isotope scoring

A0 intensity threshold (counts):

Max number of isotopes:

Isotope error threshold (ppm):

Isotope intensity threshold (counts):

Display Settings

Detail spectrum view margin (mz):

OK Cancel

OK Cancel

Figure 4. Data processing settings

(top) and color codes (bottom)

Target matching

Alpha cells (round clusters, one example marked α) are visible in the optical and selected MS images.

Results

Mass spectra were imported into the newly developed tool along with a list of targets of interest which might be expected to be detected in the sample. The settings in the tool allow to select ionization mode, adducts, mass error and intensity tolerances, and other match settings. For each target with a matching signal in the MS spectrum, the isotope pattern match, monoisotopic ion (A-number) and other characteristics were calculated and shown in a summary table.

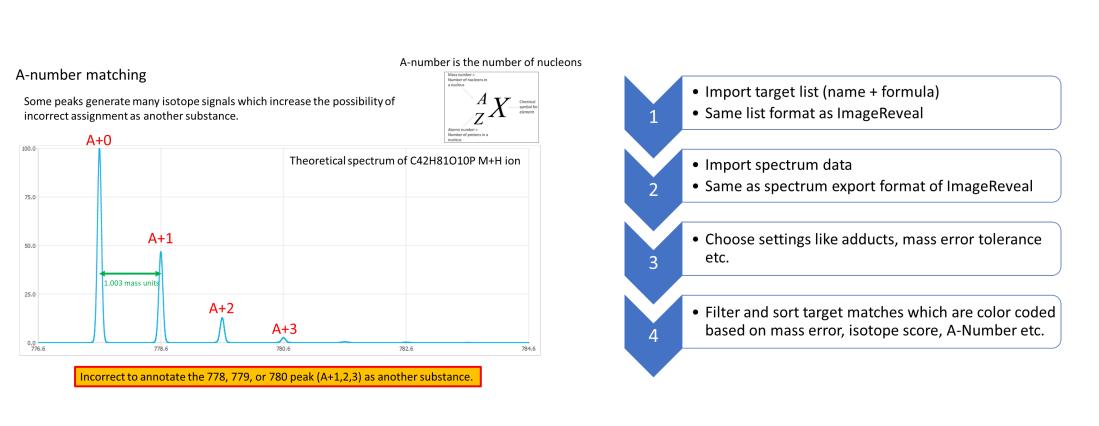


Figure 2. (Left) Concept of A-number assignment. (Right) Workflow of data processing using the tool.

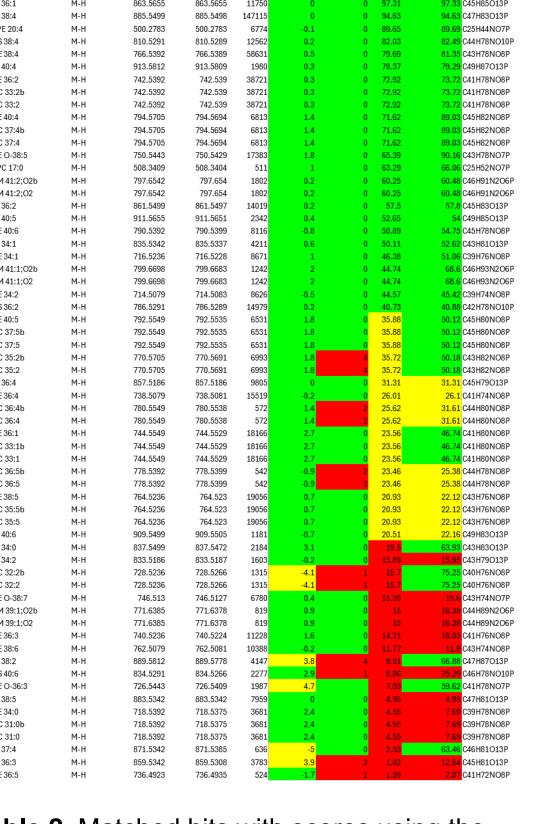


Table 2. Matched hits with scores using the newly developed tool on mouse pancreas data.

The isotope model, based on typical lipid composition and shown in figure 3, is especially important for annotation of the monoisotopic peaks in complicated spectra.

Conclusions

- Using mass accuracy and isotope pattern matching based on expected average patterns, lipid annotations could be supported or rejected.
- Combination of scores from mass accuracy, isotope pattern, and Anumber matching used to remove incorrect hits.

Acknowledgement

Tissue samples and raw data was kindly provided by Dr. Ruth Gordillo, University of Texas Southwestern Medical Center.

Disclaimer: The products and applications in this presentation are intended for Research Use Only (RUO). Not for use in diagnostic procedures.

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