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Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Leachate by EPA Method 1633A Using LC-MS/MS

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1. Overview

A reliable LC/MS/MS method for PFAS in leachate analysis according to EPA M1633A for environmental monitoring and leachate management.

2. Introduction

A recent EPA study found PFAS in over 95% of landfill leachate samples from more than 200 U.S. sites, with concentrations reaching up to 14,000 ppt across 63 different compounds (Figure 1). Despite increasing concern, there are currently no federal regulations for PFAS in landfill leachate, although some states have implemented monitoring and treatment requirements. These findings highlight the



Figure 1. Leachate Generation

urgent need for national effluent guidelines and pretreatment standards. With PFAS recognized as a persistent global contaminant, regulatory momentum is growing. EPA Method 1633A is currently the only validated multi-lab method for PFAS detection in leachate. Our laboratory has developed a sensitive and reliable LC-MS/MS method to support environmental monitoring and improve leachate management strategies.



Mobile phases consisted of 2 mM ammonium acetate in water and acetonitrile at a flow rate of 0.4 mL/min. A Shimadzu Nexcol PFAS Delay column (50 × 3.0 mm, 5 μm) was used to reduce system-derived PFAS background. Detection was performed on a Shimadzu LCMS-8060 triple quadrupole mass spectrometer using multiple reaction monitoring (MRM). Calibration ranged from 0.8 to 250 ng/mL for PFBA. LCS and spiked leachate samples were fortified at 10 ng/mL (midpoint) and 1.6 ng/mL (2× low calibrator) PFBA equivalents. Data were acquired and processed using LabSolutions and LabSolutions Insight software.

Farget Analyte N

Perfluoro-3-methox Perfluoro-4-methox Nonafluoro-3,6-dio

4. Results

Figure 4 illustrates the batch structure used to assess method robustness and instrument reliability. The batch began with a triplicate calibration curve to ensure accuracy and reproducibility. After completing the calibration, we performed a continuing calibration check throughout the batch to evaluate consistency and stability throughout the 154-injection sequence, including solvent blanks, standards, controls, and leachate samples over 37 hours of analysis.

3. Method

A composite leachate sample from the southern U.S., along with method blanks, two laboratory control samples (LCS), and two spiked leachate samples, was extracted and analyzed following EPA Method 1633A (Figure 2).

Samples were prepared using solid phase extraction (SPE) with EVOLUTE® EXPRESS WAX 150 mg/6 mL cartridges. A 100 mL volume was spiked with extracted internal standards (EIS) and passed through the cartridge, then eluted with 1% methanolic ammonium hydroxide, followed by a carbon cleanup step. Last, the extracts were spiked non-extracted internal standards (NIS).

Chromatographic separation of 40 PFAS analytes and 31 internal standards was achieved within 14 minutes using a Shim-pack Scepter C18-120 column (50 × 2.1 mm, 3 µm) with gradient elution (Figure 3, Table 1).



Figure 2. Sample preparation procedure using EPA 1633A for Leachate.

Figure 3. Chromatogram of EPA 1633A method, with %B.

Table 1. Compounds in the EPA 1633A method.

Target Analyte Name Abbrevia	tion	Target Analyte Name Abbreviation	
Perfluoroalkyl carboxylic acids		Ether sulfonic acids	
Perfluorobutanoic acid	PFBA	11-Chloroeicosafluoro-3-oxaundecane-1-sulfonic acid	11Cl-PF3OUdS
Perfluoropentanoic acid	PFPeA	9-Chlorohexadecafluoro-3-oxanonane-1-sulfonic acid	9CI-PF3ONS
Perfluorohexanoic acid	PFHxA	Perfluoro(2-ethoxyethane)sulfonic acid	PFEESA
Perfluoroheptanoic acid	PFHpA	Fluorotelomer carboxylic acids	
Perfluorooctanoic acid	PFOA	3:3 Fluorotelomer carboxylic acid	3:3 FTCA/ FPrPa
Perfluorononanoic acid	PFNA	5:3 Fluorotelomer carboxylic acid	5:3 FTCA/ FPePA
Perfluorodecanoic acid	PFDA	2H,2H,3H,3H-Perfluorodecanoic acid	7:3 FTCA/ FHpPA
Perfluoroundecanoic acid	PFUnA	Extracted Internal Standards	
Perfluorododecanoic acid	PFDoA/ PFDoDA	Perfluoro-n-[13C4]butanoic acid	13C4-PFBA
Perfluorotridecanoic acid	PFTrDA	Perfluoro-n-[13C5]pentanoic acid	13C5-PFPeA
Perfluorotetradecanoic acid	PFTeDA/ PFTreA/ PFTA	Perfluoro-n-[1,2,3,4,6-13C5]hexanoic acid	13C5-PFHxA
Perfluoroalkyl sulfonic acid	S	Perfluoro-n-[1,2,3,4-13C4]heptanoic acid	13C4-PFHpA
Perfluorobutane sulfonic acid	PFBS	Perfluoro-n-[13C8]octanoic acid	13C8-PFOA
Perfluoropentanesulfonic acid	PFPeS	Perfluoro-n-[13C9]nonanoic acid	13C9-PFNA
Perfluorohexanesulfonic acid	PFHxS	Perfluoro-n-[1,2,3,4,5,6-13C6]decanoic acid	13C6-PFDA
Perfluoroheptanesulfonic acid	PFHpS	Perfluoro-n-[1,2,3,4,5,6,7-13C7]undecanoic acid	13C7-PFUnA
Perfluorooctanesulfonic acid	PFOS	Perfluoro-n-[1,2-13C2]dodecanoic acid	13C2-PFDoA
Perfluorononanesulfonic acid	PFNS	Perfluoro-n-[1,2-13C2]tetradecanoic acid	13C2-PFTeDA
Perfluorododecanesulfonic acid	PFDS	Perfluoro-1-[2,3,4-13C3]butanesulfonic acid	13C3-PFBS
Perfluorododecanesulfonic acid	PFDoS	Perfluoro-1-[1,2,3-13C3]hexanesulfonic acid	13C3-PFHxS
Fluorotelomer sulfonic acids		Perfluoro-1-[13C8]octanesulfonic acid	13C8-PFOS
1H,1H, 2H, 2H-Perfluorohexane sulfonic acid	4:2FTS	Perfluoro-1-[13C8]octanesulfonamide	13C8-PFOSA
1H,1H, 2H, 2H-Perfluorooctane sulfonic acid	6:2FTS	N-methyl-d3-perfluoro-1-octanesulfonamidoacetic acid	D3-NMeFOSAA
1H,1H, 2H, 2H-Perfluorodecane sulfonic acid	8:2FTS	N-ethyl-d5-perfluoro-1-octanesulfonamidoacetic acid	D5-NEtFOSAA
Perfluorooctane sulfonamides		1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]hexane sulfonic acid	13C2-4:2FTS
Perfluoro-1-octanesulfonamide	PFOSA/ FOSA	1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]octane sulfonic acid	13C2-6:2FTS
N-Methylperfluorooctanesulfonamide	NMeFOSA/ MeFOSA	1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]decane sulfonic acid	13C2-8:2FTS
N-ethylperfluorooctanesulfonamide	NEtFOSA/ EtFOSA	Tetrafluoro-2-heptafluoropropoxy-13C3-propanoic acid	13C3-HFPO-DA
Perfluorooctane sulfonamidoacetic acids		N-methyl-D7-perfluorooctanesulfonamidoethanol	D7-NMeFOSE
N-methyl perfluorooctanesulfonamidoacetic acid	NMeFOSAA	N-ethyl-D9-perfluorooctanesulfonamidoethanol	D9-NEtFOSE
N-ethyl perfluorooctanesulfonamidoacetic acid	NEtFOSAA	N-ethyl-D5-perfluoro-1-octanesulfonamide	D5-NEtFOSA
Perfluorooctane sulfonamide ethanols		N-methyl-D3-perfluoro-1-octanesulfonamide	D3-NMeFOSA
2-(N-methylperfluoro-1-octanesulfonamido)-ethanol N-MeFOSE		Non-Extracted Internal Standards	
2-(N-ethylperfluoro-1-octanesulfonamido)-ethanol	N-EtFOSE	Perfluoro-n-[2,3,4-13C3]butanoic acid	13C3-PFBA
Per- and Polyfluoroether carboxylic acids		Perfluoro-n-[1,2,3,4-13C4]octanoic acid	13C4-PFOA
Hexafluoropropylene oxide dimer acid	HFPO-DA/ GenX	Perfluoro-n-[1,2-13C2]decanoic acid	13C2-PFDA
4,8-Dioxa-3H-perfluorononanoic acid	ADONA	Perfluoro-n-[1,2,3,4-13C4]octanesulfonic acid	13C4-PFOS
Perfluoro-3-methoxypropanoic acid	PFMPA	Perfluoro-n-[1,2,3,4,5-13C5] nonanoic acid	13C5-PFNA
Perfluoro-4-methoxybutanoic acid	PFMBA	Perfluoro-n-[1,2-13C2]hexanoic acid	13C2-PFHxA
Nonafluoro-3,6-dioxaheptanoic acid	NFDHA		

All 40 PFAS analytes demonstrated excellent linearity and precision, with relative standard errors (RSE) below 20% using five replicates. CCV recoveries consistently ranged between 70-130%, and concentration variability remained under 15% RSD for all analytes in the panel and under 6% for PFBA represented in Figure 4. This data confirms method and system durability despite numerous complex leachate matrix injections under high-throughput conditions.

Quantifiable PFAS were not detected in the method blank, highlighting the effectiveness of the PFAS delay column and system cleanliness in eliminating background contamination. LCS samples confirmed quantitation accuracy for the extraction process, with all native PFAS recoveries within the 70–130% range, in compliance with EPA Method 1633A criteria.



Figure 4. The batch structure designed to assess the performance and reliability. Data for PFAB CCV illustrates reproducibility with a percent RSD less than 6% for PFAB as well as good accuracy and peak shape. The CCV was less than 15% for all analytes included in the panel.

Source saturation was ruled out as a factor for low extracted internal standard (EIS) recovery based on post-extraction dilution experiments. Improved EIS recovery was observed when leachate samples were diluted *prior* to solid phase extraction (Figure 5).

WP 105

Native PFAS compounds detected in the unspiked, undiluted leachate included: 5:3 FTCA, 6:2 FTS, 7:3 FTCA, NEtFOSAA, NMeFOSAA, PFBA, PFBS, PFHpA, PFHxA, PFHxS, PFNA, PFOA, PFOS, and PFPeA.



Figure 5. Illustrates the effect of diluting the leachate samples post- and pre-extraction to improve EIS recovery. Leachate samples were diluted 2x and 5x after extraction to evaluate source saturation. In addition, leachate samples were diluted 2x and 5x before extraction to assess the SPE saturation. Undiluted leachate and diluted leachate samples were then compared to the acceptable percent recovery range per EPA 1633a for the extracted internal standard (EIS) in leachate.

5. Conclusion

The Shimadzu LCMS-8060 triple quadrupole system provided robust, stable performance over 154 injections and 37 hours of continuous PFAS analysis. With effective background reduction and rapid separation, the method follows EPA 1633A and is ideal for routine monitoring of complex leachate matrices.

6. Reference

- Tissue Samples by LC-MS/MS . Available at: compliant.pdf

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1. U.S. Environmental Protection Agency. Effluent Guidelines Program Plan 15. January 2023. Landfills Effluent Guidelines. Available at: <u>https://www.epa.gov/eg/landfills-effluent-guidelines</u> 2. U.S. Environmental Protection Agency. **Method 1633, Revision A.** December 2024. *Analysis of* Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, and https://www.epa.gov/system/files/documents/2024-12/method-1633a-december-5-2024-508-