



GCMS-QP2010 Series

**Determination of Drugs of Abuse
in Oral Fluids**

**Shimadzu
Gas Chromatograph/Mass Spectrometer**



Shimadzu Scientific Instruments (SSI) is the American subsidiary of Shimadzu Corporation, headquartered in Kyoto, Japan. Founded in 1875, Shimadzu is a \$2 billion multinational corporation with three major divisions: Medical Diagnostics, Aerospace/Industrial, and Analytical Instruments. The Analytical Division is one of the world's largest manufacturers of analytical instrumentation and environmental monitoring equipment. In addition to Japan, Shimadzu products are manufactured in China, the Philippines, the U.K., and in Portland, Oregon. International sales, service, and technical support facilities are located in Germany, Singapore, Australia, China, Brazil, and Turkey, with numerous regional offices and distributors worldwide.

In 1975, SSI corporate headquarters was established in Columbia, Maryland to provide analytical solutions to a wide range of laboratories in North, Central, and parts of South America. In the U.S., SSI has a network of more than 50 locations providing local and regional sales, service, and technical support.

Customer Training and Education Center

The Shimadzu Customer Training and Education Center (CTEC) serves as the primary educational facility for our customers. State-of-the-art interactive teaching media is employed in classroom sessions, and highly qualified instructors train customers in the operation and maintenance of their Shimadzu instruments. Course development is based on both theoretical and practical applications. We welcome and encourage colleague discussions and provide the opportunity to interact with Shimadzu product specialists.

To meet the increasing demand for more localized availability, Shimadzu offers convenient on-site training. Interactive PC-based programs have also been developed for selected courses. For more information and/or to register call 1-800-477-1227, or visit www.ssi.shimadzu.com/training.

Product Service and Support

The goal of the Shimadzu product support staff is to ensure our customers' success with their instruments. Highly trained field service technicians, strategically located throughout the country, are equipped to enable fast and efficient response to any situation. They are supported by experienced product engineers and applications specialists at the Shimadzu Technical Support Center in Columbia, MD.

Free technical support is available by contacting our Customer Service Center at 1-800-477-1227.

Your chromatography work touches peoples’ lives every day. From drug screens to criminal evidence to post-mortem analy-ses, your results are more than just numbers on paper; they can forever alter the course of a person’s life. Fast, accurate, verifiable data is everything. For that, Shimadzu’s GCMS-QP2010 Series gas chromatograph mass spectrometers provide unsurpassed performance, accuracy, speed and data reporting – all at a favorable cost/performance ratio to meet every lab’s needs.

The methods in this booklet are a compilation of work collected from working forensics laboratories. Each of these labs is a Shimadzu customer and all of the data was generated using the GCMS-QP2010 series of instruments.

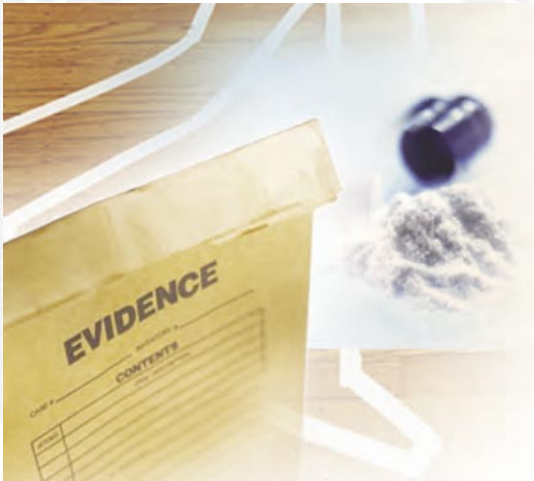


Table of Contents

Opiates	4
Cocaine and Benzoylecgonine	10
Phencyclidine	16
Amphetamines	19
THC	24
Methadone	27
About the GCMS-QP2010 Series	30
Office Locations	31

A. Opiates

Recommended Federal Cut-off: Morphine, codeine: 40 ng/mL;
6-acetylmorphine: 4 ng/mL

i. Extraction Procedure

1. From Quantisal™ device, remove 1 mL of oral fluid + buffer
2. Add 100 µL of deuterated internal standard to the calibrator and controls

Standards:

- a) D₃-codeine, D₃-morphine; D₃-6-AM at a concentration of 200 ng/mL
b) Codeine, morphine; 6-AM at a concentration of 200 ng/mL

100 μ L in 1 mL of oral fluid sample gives an internal standard concentration of 20 ng/mL (= 80 ng/mL without buffer)

Note: Acetonitrile (ACN) is a better storage solvent than methanol due to stability issues with 6-AM

Calibration Curve:

- i. Negative: 100 μ L of deuterated stock solution (200 ng/mL)
- ii. 2 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
25 μ L of 20 ng/mL stock solution (dilute 200 ng/mL 1:10)
- iii. 4 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
5 μ L of 200 ng/mL stock solution
- iv. 10 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
12.5 μ L of 200 ng/mL stock solution
- v. 20 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
25 μ L of 200 ng/mL stock solution
- vi. 40 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
50 μ L of 200 ng/mL stock solution
- vii. 80 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
100 μ L of 200 ng/mL stock solution

3. Add 0.1 M sodium phosphate buffer (pH 6.0, 1 mL). Vortex
4. Condition solid phase extraction columns (Part # 691-0353T, SPEWare, San Pedro, CA):
 - Methanol (2 mL)
 - 0.1 M phosphate buffer (pH 6.0; 2mL)
5. Add sample and allow to drain through the column
6. Wash column with:
 - Deionized water (1 mL)
 - Acetate buffer (pH 4.2; 1 mL)
 - Methanol (1 mL)
 - Ethyl acetate (1 mL)

7. Place glass collection tubes into the sample rack and elute drugs with ethyl acetate: ammonium hydroxide (98:2 v/v, 2 mL)
8. Evaporate the sample to dryness under nitrogen

Derivatization

- Reconstitute in ethyl acetate (25 µL); add BSTFA + 1% TMCS (25 µL)
- Transfer to autosampler vials, cap and heat at 70°C/20 min
- Analyze using GC/MS

ii. Analytical Procedure

Instrument: Shimadzu GCMS-QP2010
Column: RTX-XLB (Ultra low bleed, proprietary low polarity phase)
30 m length x 0.25 mm diameter x 0.25 µm film thickness

Injection volume: 2 µL
Injection Temp: 250°C
Injection mode: Splitless
Column flow: 1.3 mL/min
Linear velocity 43.3 cm/sec
Purge flow: 3 mL/min
Total flow: 49.7 mL/min.

Oven program: 150°C for 1.5 min
ramp at 20°C/min to 290°C, hold for 3 min

Ion source temperature: 230°C
Interface temperature: 250°C
Mode of operation: Standard CI mode (positive ion)
Reagent gas: Methane
Detector gain: 0.8kV above tune

Derivative: BSTFA

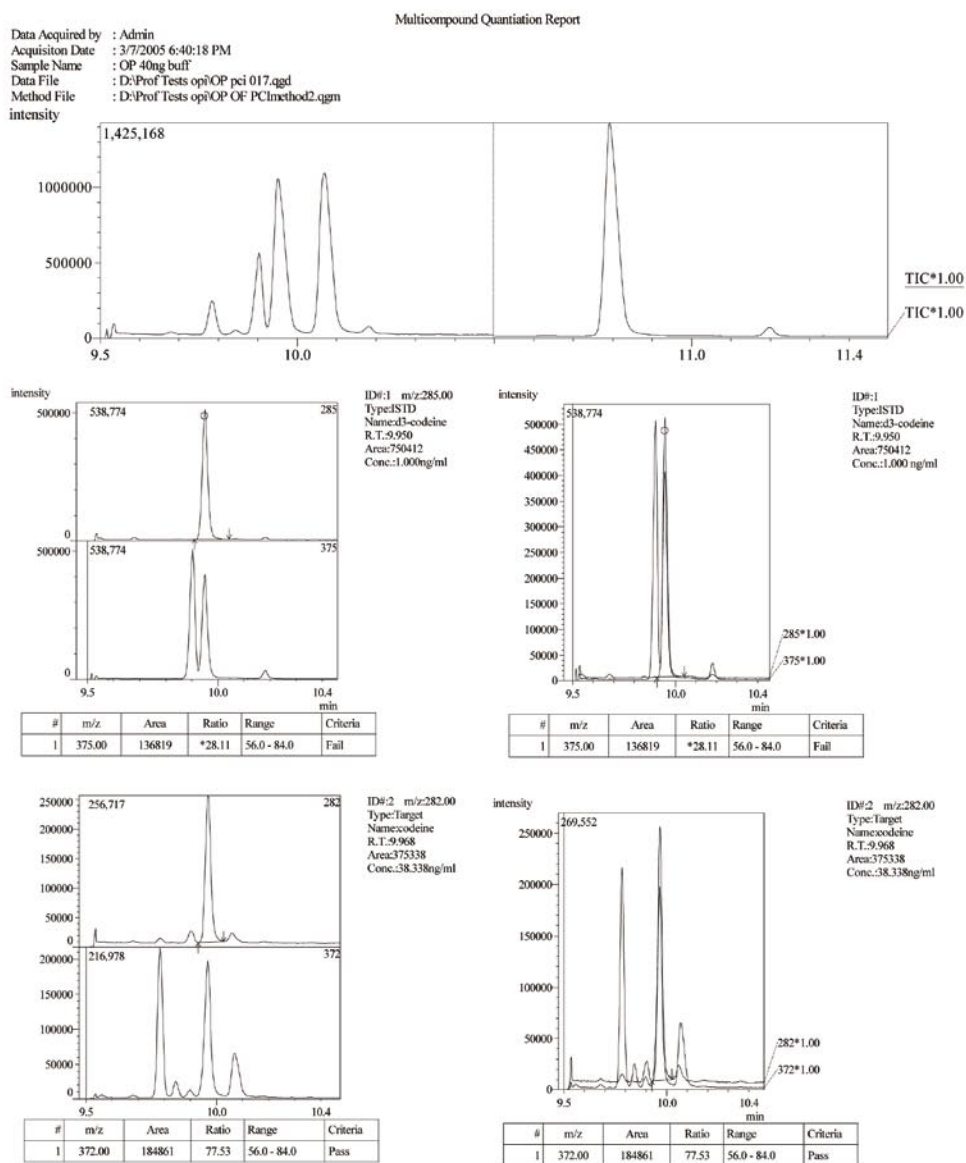
Ions monitored: 375, 285 for deuterated codeine (d3);
372, 282 for codeine;
433, 417 for deuterated morphine (d3);
430, 414 for morphine
402, 343 for deuterated 6-acetylmorphine (d3)
399, 340 for 6-acetylmorphine

In Electron Impact mode:

Ions monitored: 374.1, 237.1 for deuterated codeine (d3);
 371.1, 234.1 for codeine
 432.1, 417.1 for deuterated morphine (d3);
 429.1, 414.1 for morphine
 402.2, 343.2 for deuterated 6-acetylmorphine (d3);
 399.2, 340.2 for 6-acetylmorphine

Linearity: 0 – 80 ng/mL; limit of quantitation: 2 ng/mL

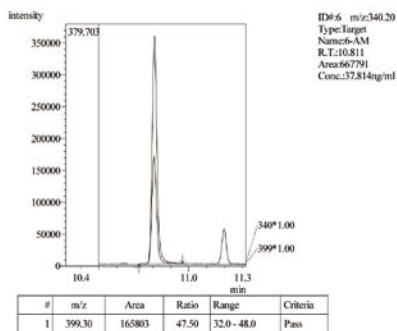
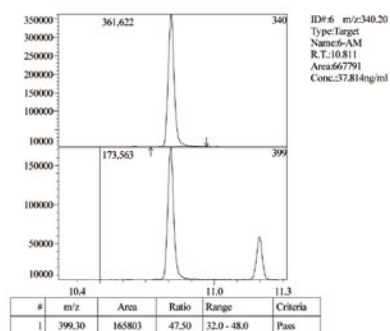
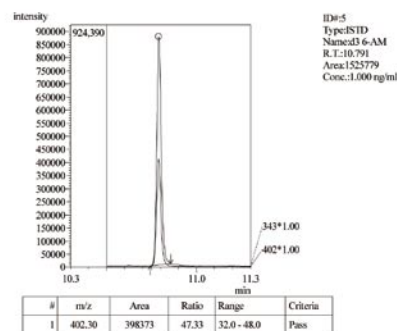
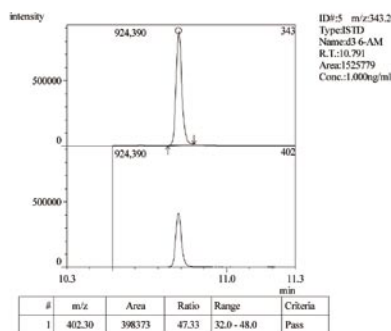
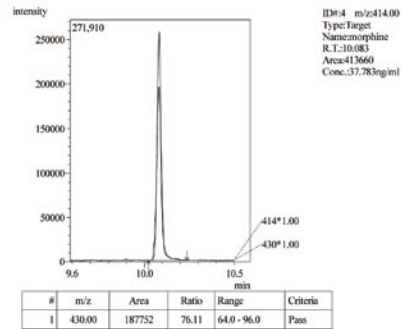
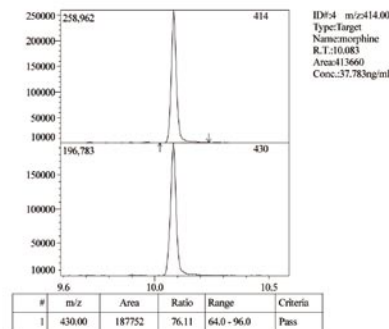
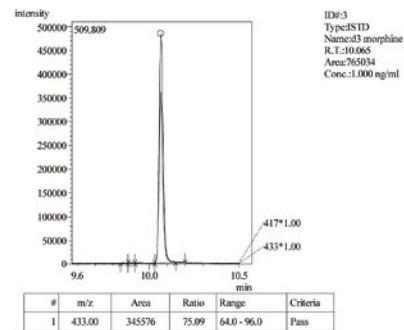
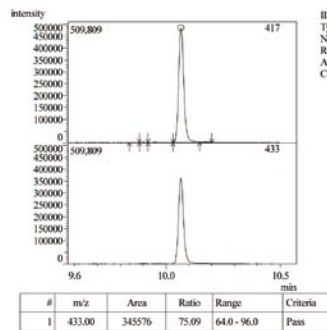
Correlation coefficients: Codeine $r^2 = 0.991$
 Morphine: $r^2 = 0.996$
 6-acetylmorphine $r^2 = 0.994$



Analyst _____ Date _____

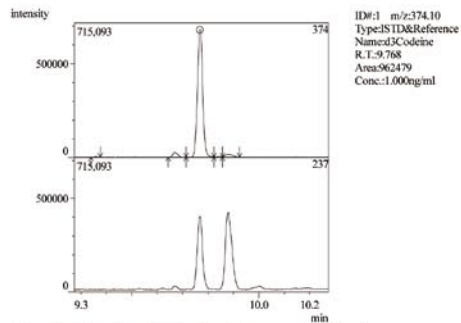
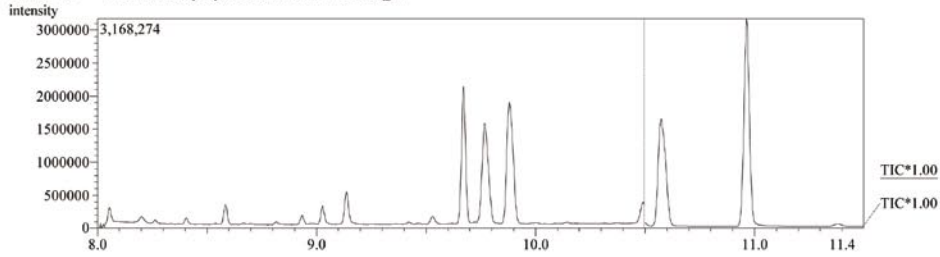
Approved by _____ Date _____

Printed on:09 Oct 2007 10:18

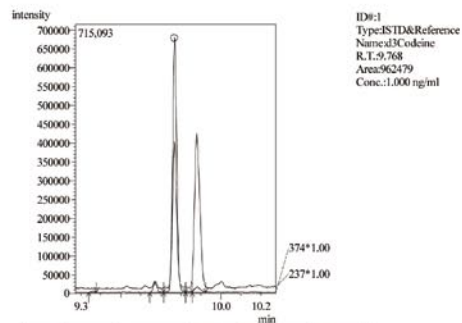


Multicomponent Quantitation Report

Data Acquired by : Admin
 Acquisition Date : 5/24/2005 12:07:00 PM
 Sample Name : OP 40ng saliva
 Data File : D:\Prof Tests opi\OP EI 047.qgd
 Method File : D:\Prof Tests opi\Opiate OF buffer curve EI 052405.qgm



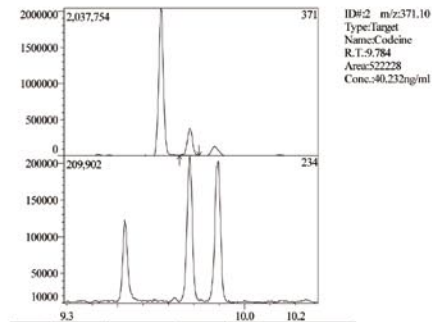
ID#1 m/z:374.10
 Type:STD&Reference
 Name:d3Codeine
 R.T.:9.768
 Area:962479
 Conc.:1.000ng/ml



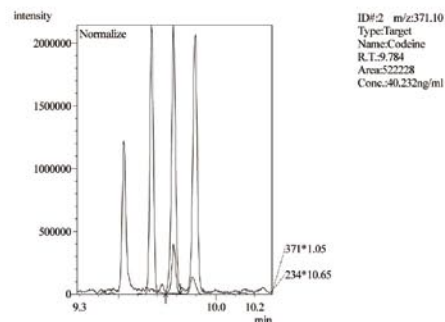
ID#1
 Type:STD&Reference
 Name:d3Codeine
 R.T.:9.768
 Area:962479
 Conc.:1.000 ng/ml

#	m/z	Area	Ratio	Range	Criteria
1	237.10	369229	57.61	30.0 - 70.0	Pass

#	m/z	Area	Ratio	Range	Criteria
1	237.10	369229	57.61	30.0 - 70.0	Pass



ID#2 m/z:371.10
 Type:Target
 Name:Codeine
 R.T.:9.784
 Area:522228
 Conc.:40.232ng/ml



ID#2 m/z:371.10
 Type:Target
 Name:Codeine
 R.T.:9.784
 Area:522228
 Conc.:40.232ng/ml

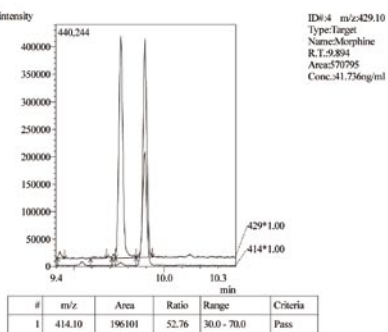
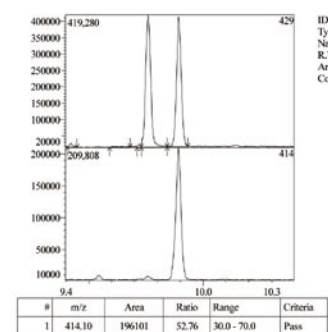
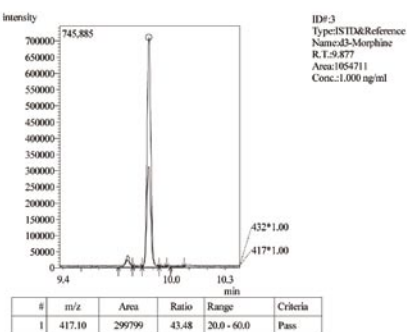
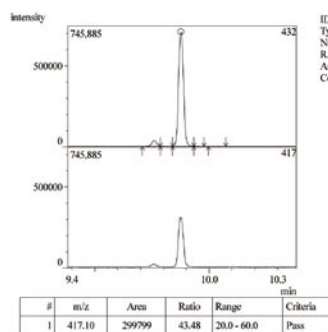
#	m/z	Area	Ratio	Range	Criteria
1	234.10	187358	53.23	30.0 - 70.0	Pass

#	m/z	Area	Ratio	Range	Criteria
1	234.10	187358	53.23	30.0 - 70.0	Pass

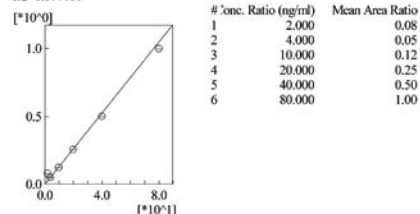
Analyst _____ Date _____

Approved by _____ Date _____

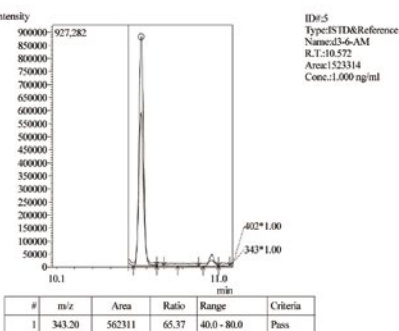
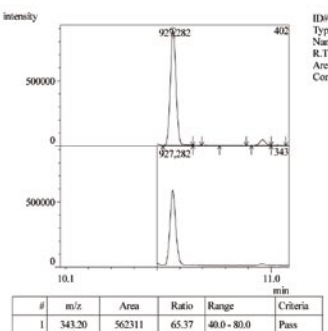
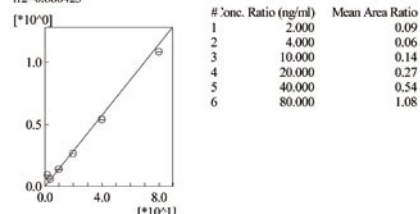
Printed on:09 Oct 2007 11:56



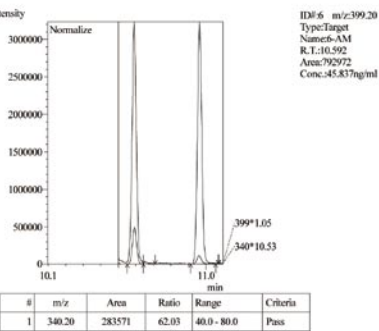
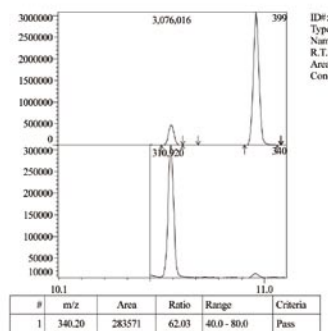
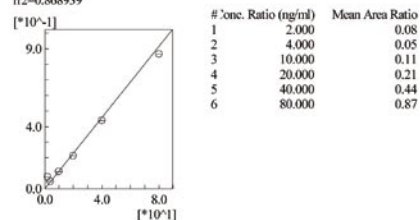
Calibration
ID#2 Mass:282.00 Name:codeine
 $f(x) = 0.013081 \cdot x + 0.000000$
 $r^2 = 0.877100$



ID#4 Mass:414.00 Name:morphine
 $f(x) = 0.014337 \cdot x + 0.000000$
 $r^2 = 0.880423$



ID#6 Mass:340.20 Name:6-AM
 $f(x) = 0.011437 \cdot x + 0.000000$
 $r^2 = 0.868939$



B. Cocaine and BZE

Recommended Federal Cut-off: Cocaine or benzoylecgonine: 8 ng/mL

i. Extraction Procedure

1. From Quantisal specimen, remove 1 mL of oral fluid + buffer
2. Add 40 μ L of deuterated internal standard to the calibrator and controls

Standards:

- c) D₃-cocaine, D₃-benzoylecgonine at a concentration of 100 ng/mL
- d) Cocaine and benzoylecgonine at a concentration of 100 ng/mL

40 μ L in 1mL of oral fluid gives an equivalent internal standard concentration of 16 ng/mL (4 ng/ml diluted)

Calibration Curve:

- i. Negative: 40 μ L of deuterated stock solution (100 ng/mL)
- ii. 4 ng/mL: 40 μ L of deuterated stock solution (100 ng/mL)
10 μ L of 100 ng/mL stock solution
- iii. 8 ng/mL: 40 μ L of deuterated stock solution (100 ng/mL)
20 μ L of 100 ng/mL stock solution
- iv. 16 ng/mL: 40 μ L of deuterated stock solution (100 ng/mL)
40 μ L of 100 ng/mL stock solution
- v. 32 ng/mL: 40 μ L of deuterated stock solution (100 ng/mL)
80 μ L of 100 ng/mL stock solution

3. Add 0.1 M potassium phosphate buffer (pH 6.0, 1 mL). Vortex
4. Condition solid phase extraction columns (Part # 691-0353T, SPEWare, San Pedro, CA):
 - Methanol (2 mL)
 - 0.1 M phosphate buffer (pH 6.0; 2mL)
5. Add sample and allow to drain through the column
6. Wash column with:
 - Deionized water (2 mL)
 - 0.1M hydrochloric acid (2 mL)
 - Methanol (3 mL)
 - Ethyl acetate (3 mL)
7. Place glass collection tubes into the sample rack and elute drugs with methylene chloride:
isopropanol: ammonium hydroxide (78:20:2 v/v. 3 mL)
8. Evaporate the sample to dryness under nitrogen

Derivatization

- Add methylene chloride (40 μ L), trifluoroethanol (20 μ L), and heptafluorobutyric anhydride (HFBA, 20 μ L) to dried extract
- Cap; allow to equilibrate for 10 minutes
- Evaporate to dryness in a vacuum oven; reconstitute in ethyl acetate (50 μ L)
- Transfer to autosampler vials for analysis using GC/MS

ii. Analytical Procedure

GC/MS: Shimadzu GCMS-QP2010

Column: RTX-XLB (Ultra low bleed, proprietary low polarity phase)
30 m length x 0.25 mm diameter x 0.25 μ m film thickness

Injection volume: 2 μ L
Injection Temp: 260°C
Injection mode: Splitless
Column flow: 1.32 mL/min
Linear velocity 43.3 cm/sec
Purge flow: 3 mL/min
Total flow: 50.4 mL/min

Oven program: 130°C for 1 min
ramped at 25°C/min to 250°C, held for 3 min
ramped at 30°C/min to 310°C

Ion source temperature: 230°C
Interface temperature: 250°C
Mode of operation: Standard CI mode (positive ion)
Reagent gas: Methane
Detector gain: 0.8kV above tune

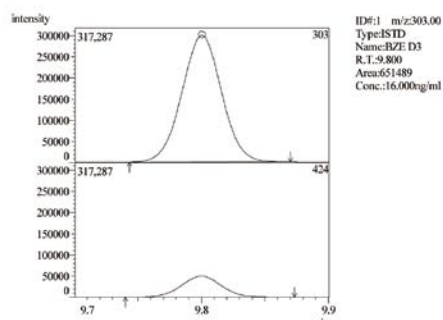
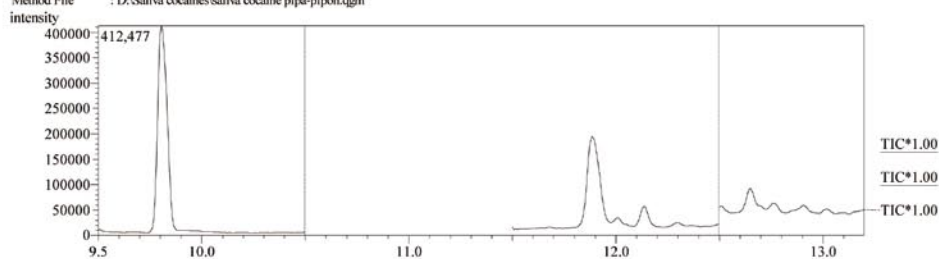
Ions monitored: 307.15, 185.15 for deuterated cocaine (d3);
304.15, 182.15 for cocaine;
375.1, 253.1 for deuterated benzoylecgonine (d3);
372.1, 250.1 for benzoylecgonine

Linearity: 0 – 32 ng/mL; Limit of quantitation: 2 ng/mL

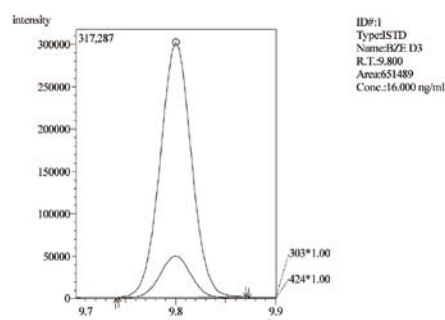
Correlation coefficients: BZE $r^2 = 0.9998$
Cocaine: $r^2 = 0.9985$

Multicompound Quantitation Report

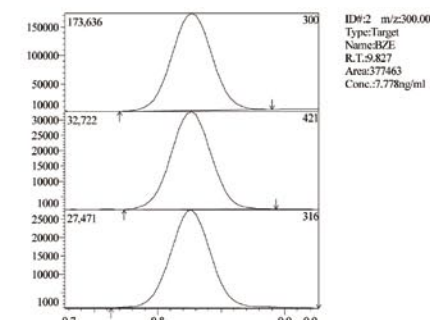
Data Acquired by : Admin
 Acquisition Date : 7/27/2005 8:38:51 PM
 Sample Name : 8ng
 Data File : D:\Saliva cocaine\coc004.qgd
 Method File : D:\Saliva cocaine\saliva cocaine pfpa-pfph.qgm



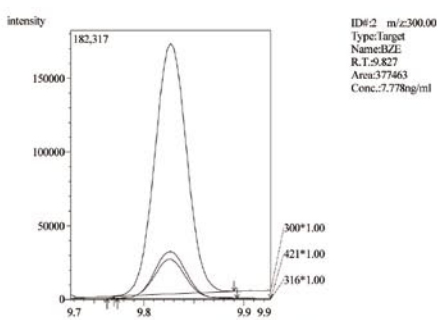
#	m/z	Area	Ratio	Range	Criteria
1	423.95	109542	16.81	14.4 - 21.6	Pass



#	m/z	Area	Ratio	Range	Criteria
1	423.95	109542	16.81	14.4 - 21.6	Pass



#	m/z	Area	Ratio	Range	Criteria
1	420.95	70785	18.75	14.0 - 26.0	Pass
2	316.00	59634	15.80	12.0 - 18.0	Pass

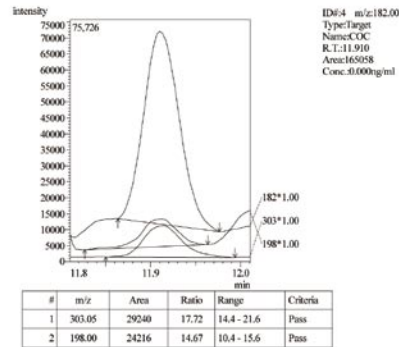
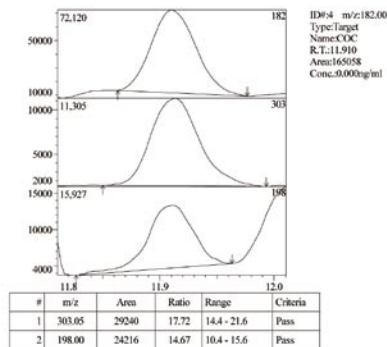
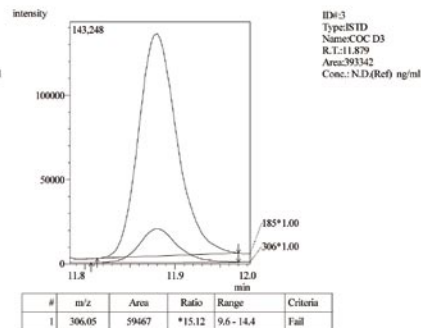
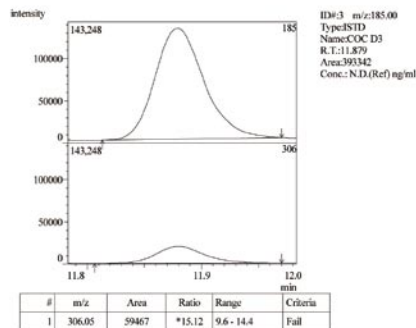


#	m/z	Area	Ratio	Range	Criteria
1	420.95	70785	18.75	14.0 - 26.0	Pass
2	316.00	59634	15.80	12.0 - 18.0	Pass

Analyst _____ Date _____

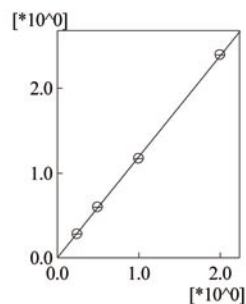
Approved by _____ Date _____

Printed on:22 Oct 2007 12:16



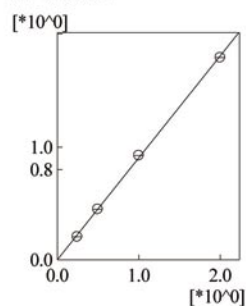
Calibration

ID#2 Mass:300.00 Name:BZE
 $f(x)=1.191832*x+0.000000$
 $r^2=0.999841$



#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	0.250	0.29
2	0.500	0.60
3	1.000	1.17
4	2.000	2.40

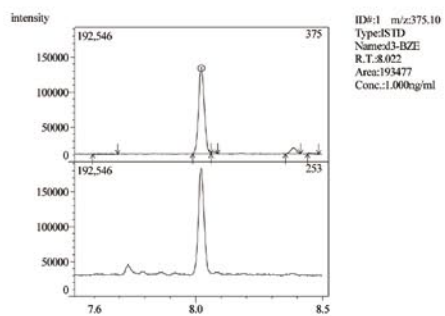
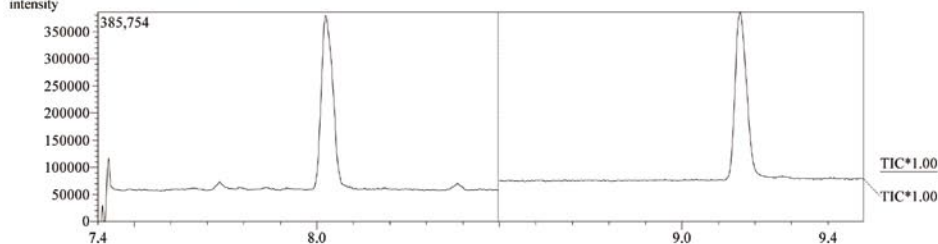
ID#4 Mass:182.00 Name:COC
 $f(x)=0.898234*x+0.000000$
 $r^2=0.999324$



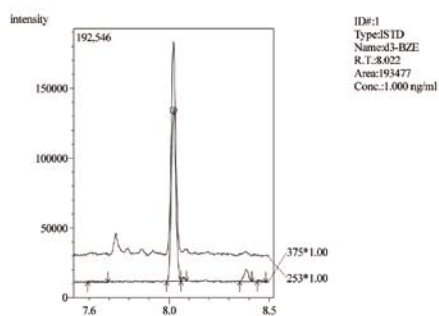
#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	0.250	0.21
2	0.500	0.45
3	1.000	0.92
4	2.000	1.79

Multicomponent Quantitation Report

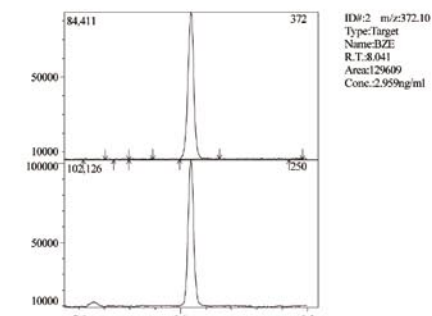
Data Acquired by : Admin
 Acquisition Date : 3/3/2005 6:28:22 PM
 Sample Name : coc 8ng/ml
 Data File : D:\cocaine saliva DT\coc 003.qgd
 Method File : D:\cocaine saliva DT\cocaine HFBA.qgm



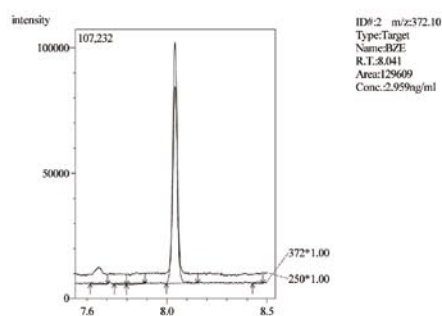
#	m/z	Area	Ratio	Range	Criteria
1	253.10	144052	121.59	96.0 - 144.0	Pass



#	m/z	Area	Ratio	Range	Criteria
1	253.10	144052	121.59	96.0 - 144.0	Pass



#	m/z	Area	Ratio	Range	Criteria
1	250.10	88836	117.72	96.0 - 144.0	Pass

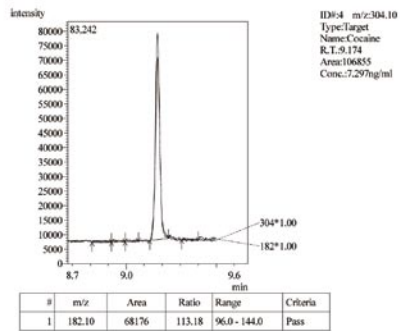
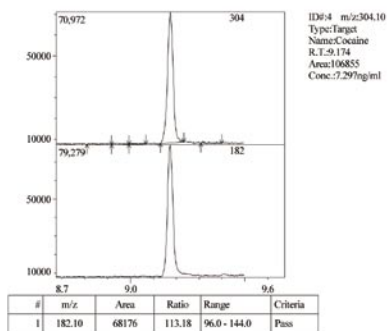
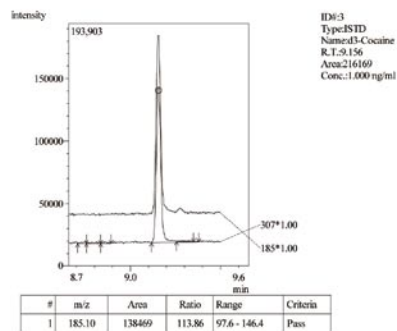
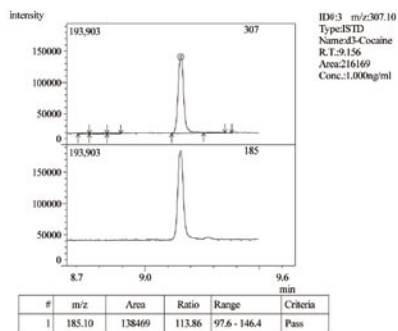


#	m/z	Area	Ratio	Range	Criteria
1	250.10	88836	117.72	96.0 - 144.0	Pass

Analyst _____ Date _____

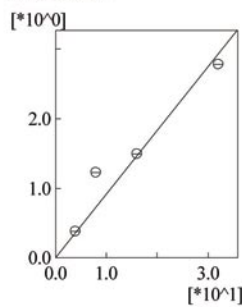
Approved by _____ Date _____

Printed on: 22 Oct 2007 12:21



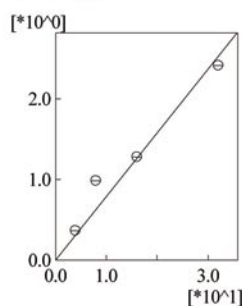
Calibration

ID#2 Mass:372.10 Name:BZE
 $f(x) = 0.091169 \cdot x + 0.000000$
 $r^2 = 0.950493$



#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	4.000	0.38
2	8.000	1.22
3	16.000	1.49
4	32.000	2.78

ID#4 Mass:304.10 Name:Cocaine
 $f(x) = 0.078733 \cdot x + 0.000000$
 $r^2 = 0.970739$



#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	4.000	0.37
2	8.000	0.99
3	16.000	1.28
4	32.000	2.41

C. Phencyclidine

Recommended Federal Cut-off: PCP: 10 ng/mL

i. Extraction Procedure

1. Measure out 1 mL of Quantisal buffer (or appropriate amount of collection buffer corresponding to 250 μ L oral fluid)
2. Add 20 μ L of deuterated internal standard to the calibrator and controls

Standards:

- a. D5-phencyclidine at a concentration of 250 ng/mL
- b. Phencyclidine at a concentration of 250 ng/mL

20 μ L in 1 mL of oral fluid gives an equivalent internal standard concentration of 20 ng/mL (5 x 4)

Calibration Curve:

- | | |
|----------------|---|
| i. Negative: | 20 μ L of deuterated stock solution (250 ng/mL) |
| ii. 5 ng/mL: | 20 μ L of deuterated stock solution (250 ng/mL)
5 μ L of 250 ng/mL stock solution |
| iii. 10 ng/mL: | 20 μ L of deuterated stock solution (250 ng/mL)
10 μ L of 250 ng/mL stock solution |
| iv. ng/mL: | 20 μ L of deuterated stock solution (250 ng/mL)
20 μ L of 250 ng/mL stock solution |
| v. ng/mL: | 20 μ L of deuterated stock solution (250 ng/mL)
40 μ L of 250 ng/mL stock solution |

3. Add 0.1M sodium bicarbonate buffer (pH 8.0, 1mL); vortex
4. Place extraction tubes (SPEWare 691-0353T) onto the vacuum manifold
5. Label columns. Condition each column:
 - Methanol (2 mL)
 - 0.1M phosphate buffer (pH 6.0, 2 mL)

Important: Do not allow the column bed to go dry.
6. Pour each sample through extraction column. Allow the sample to flow through the column. Dry.
7. Rinse each column with:
 - DI water (1 mL), dry for 1 min
 - 0.1M acetate buffer (pH 4.5, 1 mL), dry for 1 min
 - Methanol (1 mL), dry for 5 min
 - Ethyl acetate (1 mL)
8. Place labeled glass tubes into manifold; wipe the tips.
9. Elute drugs: ethyl acetate + 2% ammonium hydroxide (2 mL)
10. Evaporate to dryness under nitrogen (20 psi /37°C)
11. Reconstitute in ethyl acetate (30 μ L); Vortex.
12. Transfer to auto-sampler vials

ii. Analytical Procedure

System: Shimadzu GCMS-QP2010

Column: RTX-XLB (Ultra low bleed, proprietary low polarity phase)
30 m length x 0.25 mm diameter x 0.25 μ m film thickness

Injection volume: 2 μ L
Injection Temp: 250°C
Injection mode: Splitless
Column flow: 1 mL/min
Linear velocity 36.5 cm/sec
Purge flow: 3 mL/min
Total flow: 38.9 mL/min

Oven program: 60°C for 1 min
ramp at 25°C/min to 200°C, hold for 5.2 min
ramp at 25°C/min to 300°C

Ion source temperature: 230°C
Interface temperature: 250°C
Mode of operation: Electron Impact
Detector gain: 0.8kV above tune

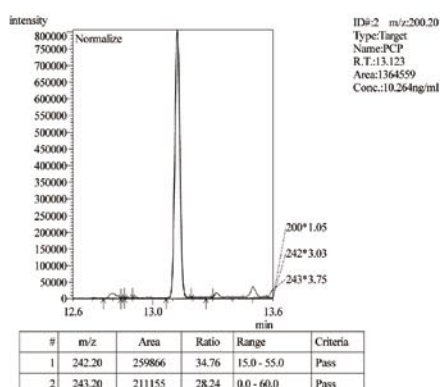
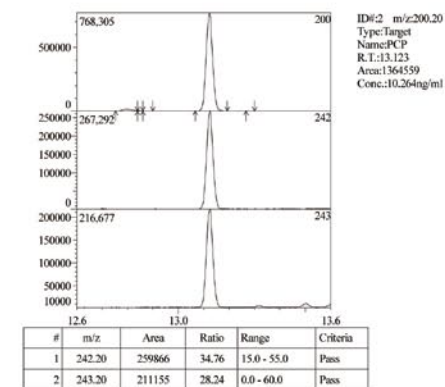
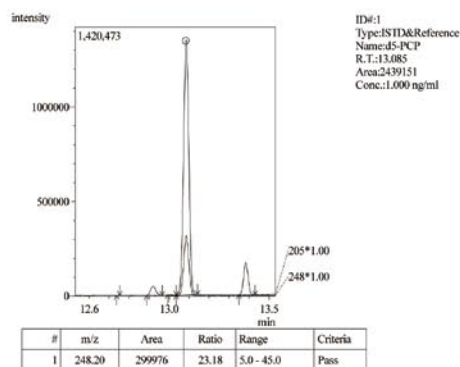
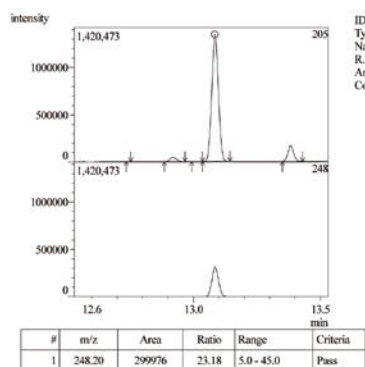
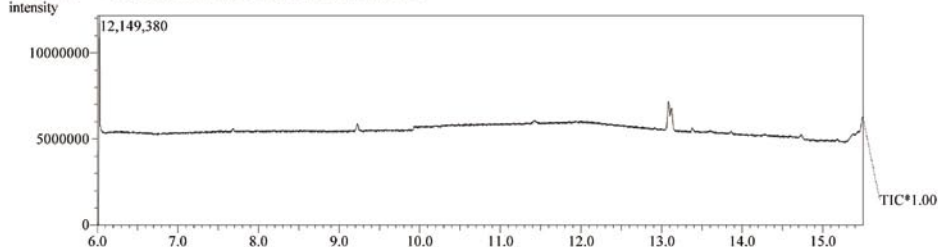
Ions monitored: 205.2, 248.2 for deuterated PCP (d5);
200.2, 243.2, 242.2 for PCP

Linearity: 0 – 40 ng/mL; Limit of quantitation: 5 ng/mL

Correlation coefficients: PCP $r^2 = 0.9998$

Data Acquired by : Admin
 Acquisition Date : 6/10/2005 12:50:54 PM
 Sample Name : 10 ng/mL_ext
 Data File : D:\PCP\PCP DATA\0101003.QGD
 Method File : C:\GCMSolution\Data\PCP\PCP Data\PCP EI Method.qgm

Multicomponent Quantitation Report



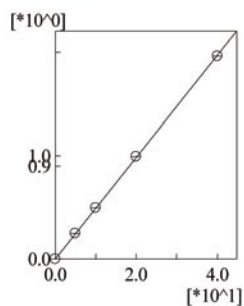
Analyst _____ Date _____

Approved by _____ Date _____

Printed on:22 Oct 2007 14:14

Calibration

ID#:2 Mass:200.20 Name:PCP
 $f(x)=0.049240*x+0.000000$
 $r^2=0.999932$



#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	0.000	0.00
2	5.000	0.25
3	10.000	0.50
4	20.000	1.00
5	40.000	1.96

D. Amphetamines

Recommended Federal Cut-off:

Methamphetamine, amphetamine, MDMA, MDA, MDEA: 50 ng/mL

Cannot report methamphetamine as positive without amphetamine present above the limit of detection

i. Extraction Procedure

1. Measure out 1 mL of Quantisal buffer (or appropriate amount of collection buffer corresponding to 250 μ L oral fluid)
2. Add 50 μ L of deuterated internal standard to the calibrator and controls

Standards:

- a) D5-amphetamine, D5-methamphetamine, D5-MDMA, D5-MDA and D5-MDEA at a concentration of 250 ng/mL
- b) Amphetamine, methamphetamine, MDMA, MDA and MDEA at a concentration of 250 ng/mL

50 μ L of (A) in 1 mL of oral fluid gives an equivalent internal standard concentration of 50 ng/mL (12.5 x 4)

Calibration Curve:

- i. Negative: 50 μ L of deuterated stock solution (250 ng/mL)
- ii. 25 ng/mL: 50 μ L of deuterated stock solution (250 ng/mL)
25 μ L of 250 ng/mL stock solution
- iii. 50 ng/mL: 50 μ L of deuterated stock solution (250 ng/mL)
50 μ L of 250 ng/mL stock solution
- iv. 100 ng/mL: 50 μ L of deuterated stock solution (250 ng/mL)
100 μ L of 250 ng/mL stock solution
- v. 200 ng/mL: 50 μ L of deuterated stock solution (250 ng/mL)
200 μ L of 250 ng/mL stock solution

3. To specimens, add 0.1M potassium phosphate buffer (pH 6.0, 1 mL); vortex
4. Place extraction tubes (SPEWare 691-0353T) onto the vacuum manifold
5. Label columns. Condition each column:
 - Methanol (2 mL)
 - 0.1M phosphate buffer (pH 6.0, 2 mL)*Important: Do not allow the column bed to go dry.*
6. Allow the sample to flow through the column. Dry.
7. Rinse each column with:
 - DI water (1mL)
 - 0.1M acetate buffer (pH 4, 1 mL)
 - Methanol (1 mL)
 - Ethyl acetate (1 mL); dry for 5 min; 30 psi
8. Place labeled glass tubes into manifold; wipe the tips.
9. Elute drugs: ethyl acetate + 2% ammonium hydroxide (2 mL)

10. Dry samples under nitrogen to dryness.
11. After 5 min add one drop of 0.35M H₂SO₄; acetone (10:90 v,v); After 10 min add another drop.
12. Add heptafluorobutyric anhydride (HFBA, 20 µL); heat at 60°C/20 min
13. Evaporate to dryness in vacuum oven
14. Reconstitute in ethyl acetate (60 µL); Vortex
15. Transfer to auto-sampler vials; analyze by GC/MS

ii. Analytical Procedure

System: Shimadzu GCMS-QP2010

Column: RTX-XLB (Ultra low bleed, proprietary low polarity phase)
30 m length x 0.25 mm diameter x 0.25 µm film thickness

Injection volume: 2 µL
Injection Temp: 150°C
Injection mode: Splitless
Column flow: 1.3 mL/min
Linear velocity 41.6 cm/sec
Purge flow: 0 mL/min
Total flow: 46.7 mL/min

Oven program: 60°C for 1 min
ramp at 25°C/min to 140°C, hold for 4 min
ramp at 30°C/min to 200°C, hold for 3 min
ramp at 40°C/min to 300°C

Ion source temperature: 220°C
Interface temperature: 250°C
Mode of operation: Electron Impact
Detector gain: 0.8kV above tune

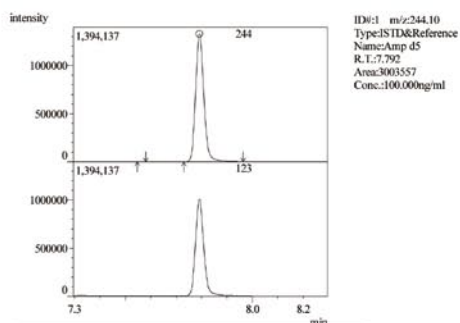
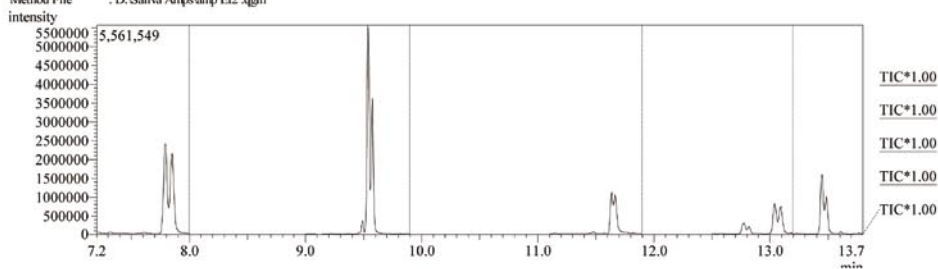
Linearity: 0 – 200 ng/mL; Limit of quantitation: 25 ng/mL
Correlation coefficients: r² = 0.9998

Acquisition Parameter File: Amphetamine Acquisition
Group Entries: Number of Groups: 5

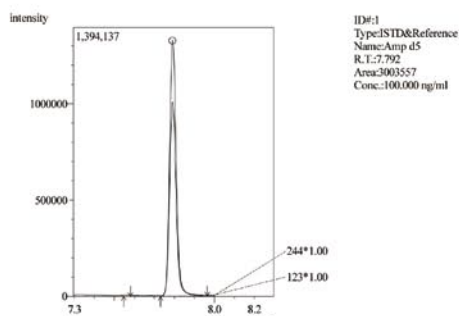
Ions	*Quantifying ion	Retention Time (min)
Group 1: 244.1*, 123.2 (d5 Amphetamine); 240.1*, 118.15, 91.1 (Amphetamine)		7.7 min
Group 2: 258.05*, 213.05 (d5 Methamphetamine); 254.1*, 210.05*, 118.15 (Methamphetamine)		9.5 min
Group 3: *136.15, 380.2 (d5 MDA); 135.15*, 162.15, 375.15 (MDA)		11.6 min
Group 4: 258.1*, 213.05 (d5 MDMA); 254.05*, 210.05, 162.15 (MDMA)		13.0 min
Group 5: *273.15, 241.1 (d5 MDEA); 268.1*, 240.1, 162.1 (MDEA)		13.4 min

Multicomponent Quantitation Report

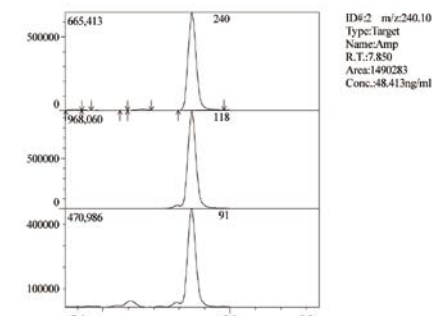
Data Acquired by : Admin
 Acquisition Date : 7/22/2005 2:24:47 PM
 Sample Name : 50ng ext saliva
 Data File : D:\Saliva Amps\Amp008.qgd
 Method File : D:\Saliva Amps\amp EI2 .agm



#	m/z	Area	Ratio	Range	Criteria
1	123.20	986579	75.20	56.0 - 84.0	Pass



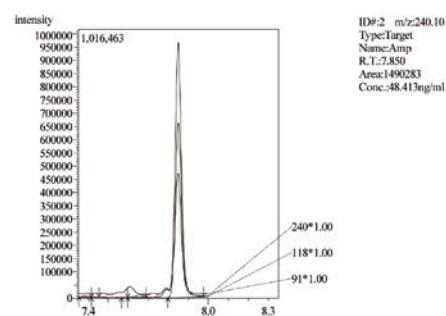
#	m/z	Area	Ratio	Range	Criteria
1	123.20	986579	75.20	56.0 - 84.0	Pass



#	m/z	Area	Ratio	Range	Criteria
1	118.15	917804	142.04	104.0 - 156.0	Pass
2	91.10	432410	66.92	48.0 - 72.0	Pass

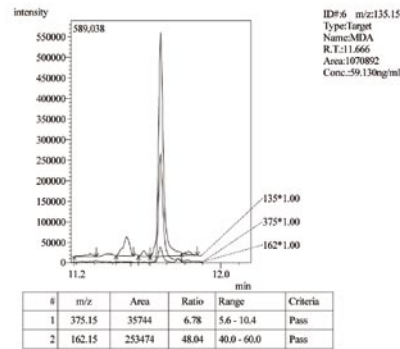
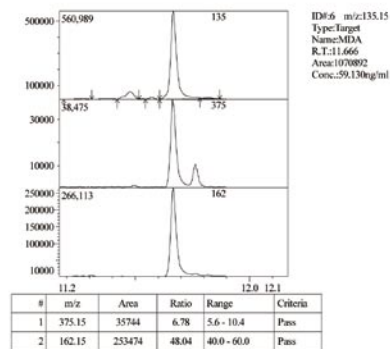
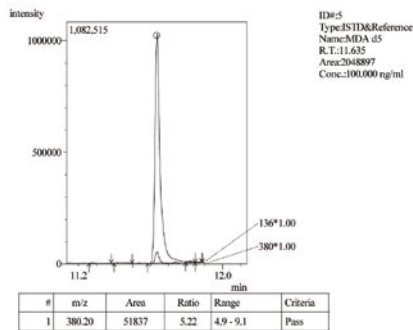
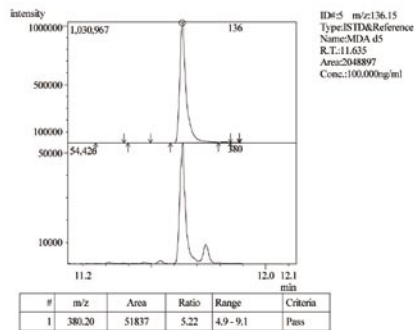
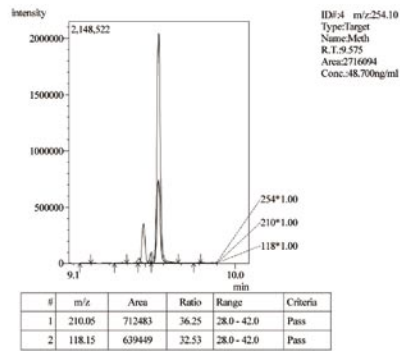
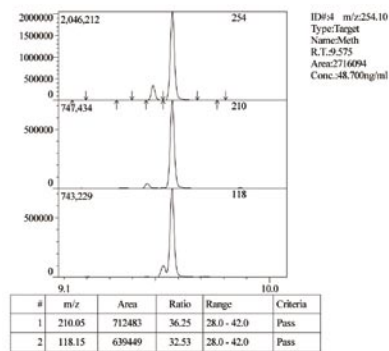
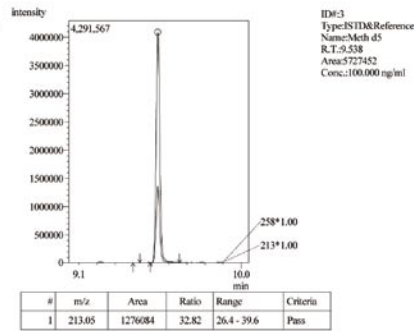
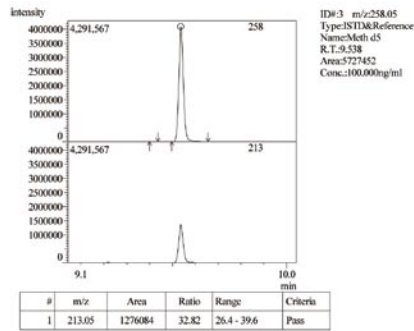
Analyst _____ Date _____

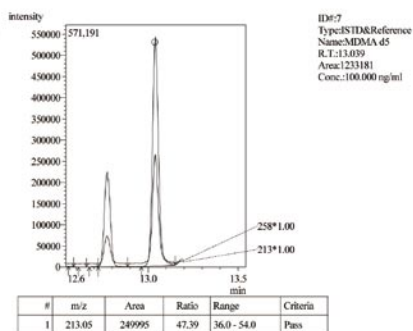
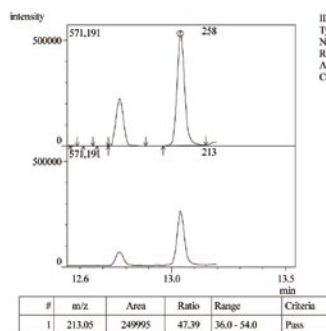
Printed on:22 Oct 2007 14:28



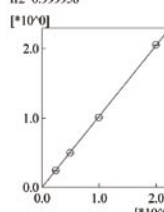
#	m/z	Area	Ratio	Range	Criteria
1	118.15	917804	142.04	104.0 - 156.0	Pass
2	91.10	432410	66.92	48.0 - 72.0	Pass

Approved by _____ Date _____

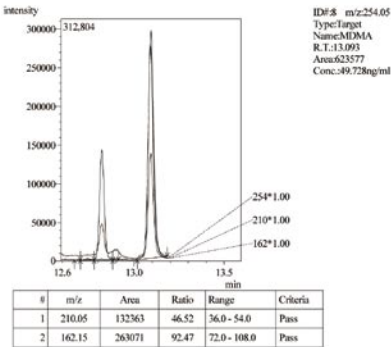
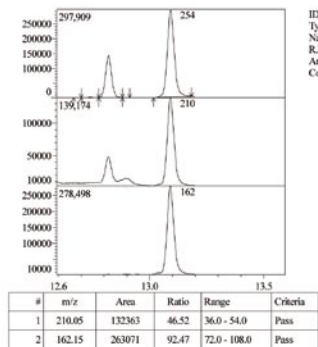




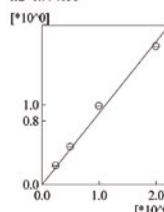
Calibration
ID#2 Mass:240.10 Name:Amp
 $f(x) = 1.020487 \times x + 0.000000$
 $r^2 = 0.999958$



#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	0.250	0.24
2	0.500	0.50
3	1.000	1.01
4	2.000	2.05

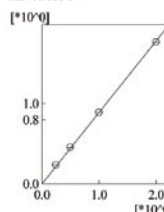


ID#4 Mass:254.10 Name:Meth
 $f(x) = 0.890418 \times x + 0.000000$
 $r^2 = 0.994056$

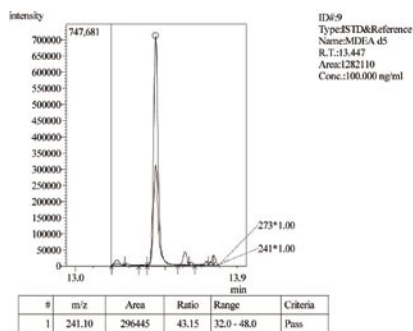
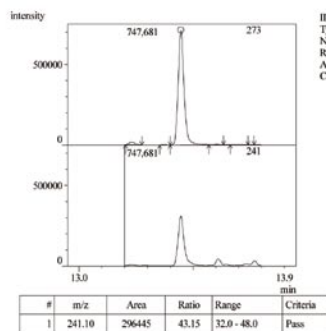


#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	0.250	0.24
2	0.500	0.47
3	1.000	0.98
4	2.000	1.73

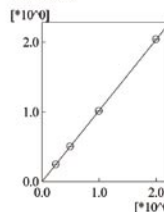
ID#6 Mass:135.15 Name:MDA
 $f(x) = 0.883944 \times x + 0.000000$
 $r^2 = 0.999984$



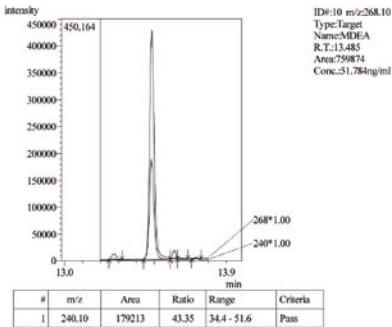
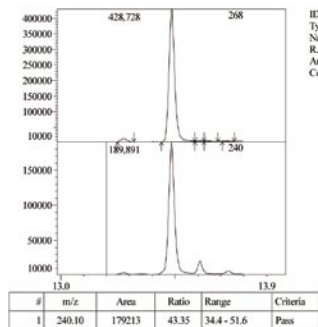
#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	0.250	0.24
2	0.500	0.45
3	1.000	0.89
4	2.000	1.76



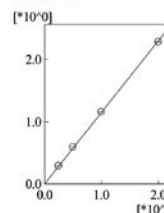
ID#8 Mass:254.05 Name:MDMA
 $f(x) = 1.016862 \times x + 0.000000$
 $r^2 = 0.999971$



#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	0.250	0.25
2	0.500	0.51
3	1.000	1.01
4	2.000	2.04



ID#10 Mass:268.10 Name:MDEA
 $f(x) = 1.144507 \times x + 0.000000$
 $r^2 = 0.999914$



#	Conc. Ratio (ng/ml)	Mean Area Ratio
1	0.250	0.29
2	0.500	0.59
3	1.000	1.16
4	2.000	2.28

E. THC

Recommended Federal Cut-off: THC 2 ng/mL

i. Extraction Procedure

1. Aliquot 1 mL of Quantisal new buffer (=0.25 mL neat oral fluid)
2. Add 20 μ L of 1000 ng/mL solution of deuterated (d3-THC) (20 ng; 80 ng/mL)

Standards:

- a) D3-THC at a concentration of 1000 ng/mL
- b) THC at a concentration of 100 ng/mL

Calibration Curve:

- i. Negative: 20 μ L of deuterated stock solution (1000 ng/mL)
- ii. 1 ng/mL: 20 μ L of deuterated stock solution (1000 ng/mL)
50 μ L of 10 ng/mL stock solution (1:10 of 100 ng/mL)
- iii. 2 ng/mL: 20 μ L of deuterated stock solution (1000 ng/mL)
100 μ L of 10 ng/mL stock solution (1:10 of 100 ng/mL)
- iv. 4 ng/mL: 20 μ L of deuterated stock solution (1000 ng/mL)
20 μ L of 100 ng/mL stock solution
- v. 8 ng/mL: 20 μ L of deuterated stock solution (1000 ng/mL)
40 μ L of 100 ng/mL stock solution

3. Add 0.1M acetate buffer (pH 4.5, 1 mL)
4. Condition SPEWare columns:
 - Methanol (0.5 mL),
 - 0.1M acetic acid (100 μ L)
5. Pour sample into column and pass through at a flow rate of 1ml / min
6. Wash column 80 : 20 D.I. H₂O : acetic acid (1ml)
40 : 60 D.I. H₂O: methanol (1ml)
7. Dry column (5 min; 30 psi)
8. Elute samples: hexane: glacial acetic acid (98:2, 0.8 mL)
9. Evaporate sample to dryness
10. Add ethyl acetate (50 μ L); transfer into auto sampler vial
11. Add BSTFA (20 μ L); heat at 60° C /15 min

ii. Analytical Procedure

System: Shimadzu GCMS-QP2010

Column: RTX-XLB (Ultra low bleed, proprietary low polarity phase)
30 m length x 0.25 mm diameter x 0.25 µm film thickness

Injection volume: 2 µL
Injection Temp: 250°C
Injection mode: Splitless
Column flow: 1.39 mL/min
Linear velocity: 44.4 cm/sec
Total flow: 50 mL/min
Oven program: 125°C for 0.2 min
ramp at 20°C/min to 250°C, hold for 3 min
ramp at 30°C/min to 300°C

Ion source temperature: 220°C
Interface temperature: 280°C
Mode of operation: Electron impact
Detector gain: 0.8kV above tune

Ions monitored: 389, 374 for deuterated THC (d3);
386, 371 for THC

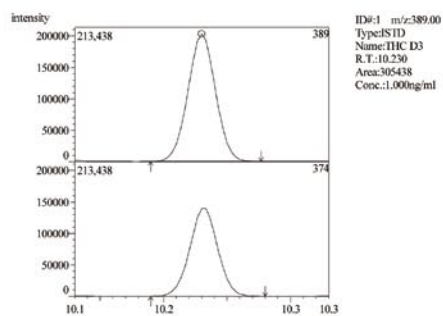
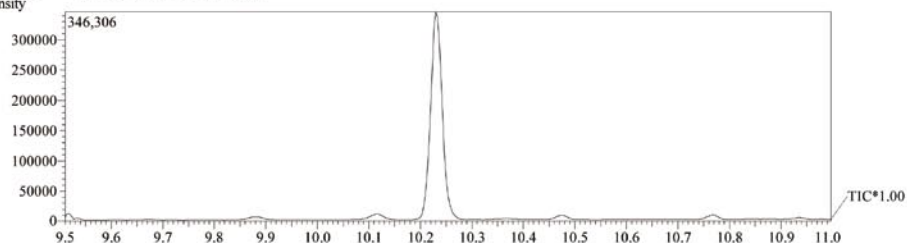
The limit of quantitation of the method was 1 ng/mL

Reference

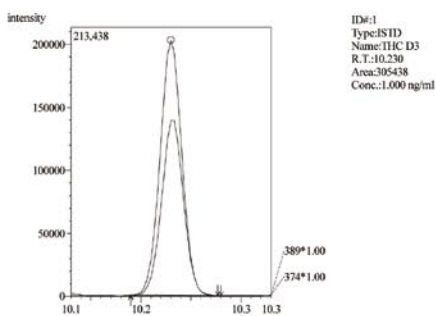
<http://www.shimadzu.com/apps/appnotes/GCMS%20QP-2010%20THC%20Saliva.pdf>

Data Acquired by : Admin
 Acquisition Date : 2/17/2005 4:18:34 PM
 Sample Name : 2 ng cal
 Data File : D:\Oral fluid THC\THC03.qgd
 Method File : D:\Oral fluid THC\THCOFDT.qgm
 intensity

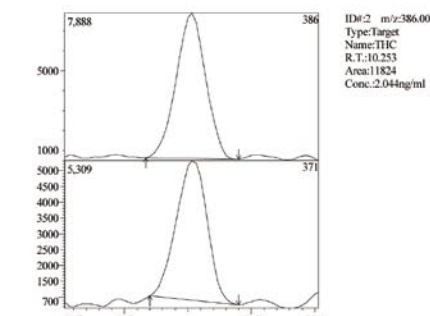
Multicomponent Quantitation Report



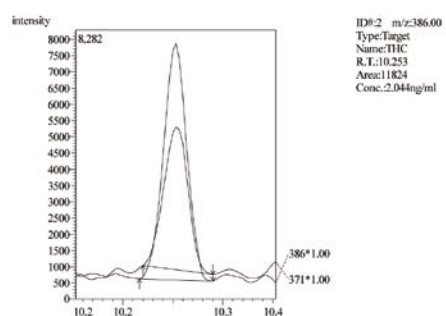
#	m/z	Area	Ratio	Range	Criteria
1	374.00	213028	69.75	55.2 - 82.8	Pass



#	m/z	Area	Ratio	Range	Criteria
1	374.00	213028	69.75	55.2 - 82.8	Pass



#	m/z	Area	Ratio	Range	Criteria
1	371.00	7448	62.99	48.3 - 89.7	Pass



#	m/z	Area	Ratio	Range	Criteria
1	371.00	7448	62.99	48.3 - 89.7	Pass

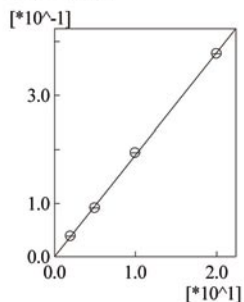
Analyst _____ Date _____

Approved by _____ Date _____

Printed on:22 Oct 2007 14:42

Calibration

ID#2 Mass:386.00 Name:THC
 $f(x) = 0.018938 * x + 0.000000$
 $r^2 = 0.999598$



#	Conc. Ratio (ng/ml)	Mean Area Ratio
2	2.000	0.04
3	5.000	0.09
4	10.000	0.19
5	20.000	0.38

F. Methadone

i. Extraction Procedure

1. From Quantisal specimen, remove 1 mL of oral fluid + buffer
2. Add 100 μ L of deuterated internal standard to the calibrator and controls

Standards:

- a) D9-methadone at a concentration of 200 ng/mL
- b) Methadone at a concentration of 200 ng/mL

100 μ L in 1 mL of oral fluid sample gives an internal standard concentration of 20 ng/mL (= 80 ng/mL neat oral fluid)

Calibration Curve:

- i. Negative: 100 μ L of deuterated stock solution (200 ng/mL)
- ii. 10 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
12.5 μ L of 200 ng/mL stock solution
- iii. 20 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
25 μ L of 200 ng/mL stock solution
- iv. 40 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
50 μ L of 200 ng/mL stock solution
- v. 80 ng/mL: 100 μ L of deuterated stock solution (200 ng/mL)
100 μ L of 200 ng/mL stock solution

3. Add 0.1 M sodium bicarbonate buffer (pH 8.0, 1 mL). Vortex
4. Condition solid phase extraction columns (Part # 691-0353T, SPEWare, San Pedro, CA):
 - Methanol (2 mL)
 - 0.1 M phosphate buffer (pH 6.0; 2mL)
5. Add sample and allow to drain through the column
6. Wash column with:
 - Deionized water (1 mL)
 - 0.1 M Acetate buffer (pH 4.2; 1 mL)
 - Methanol (1 mL)
 - Ethyl acetate (1 mL); dry for 2 min
7. Place glass collection tubes into the sample rack
8. Elute drugs with ethyl acetate: ammonium hydroxide (98:2 v/v, 2 mL)
9. Evaporate the sample to dryness under nitrogen
10. Reconstitute in ethyl acetate (40 μ L); Transfer to autosampler vials
11. Analyze using GC/MS

ii. Analytical Procedure

System: Shimadzu GCMS-QP2010

Column: RTX-XLB (Ultra low bleed, proprietary low polarity phase)
30 m length x 0.25 mm diameter x 0.25 µm film thickness

Injection volume: 2 µL
Injection Temp: 250°C
Injection mode: Splitless
Column flow: 1.2 mL/min
Linear velocity: 40.8 cm/sec
Total flow: 46.4 mL/min

Oven program: 110°C for 1 min
ramp at 30°C/min to 290°C, hold for 1.5 min

Ion source temperature: 230°C
Interface temperature: 300°C
Mode of operation: Standard electron impact (EI) mode
Detector gain: 0.8kV above tune

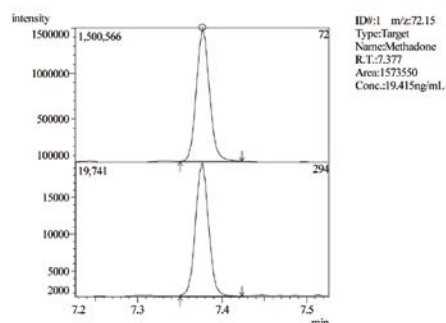
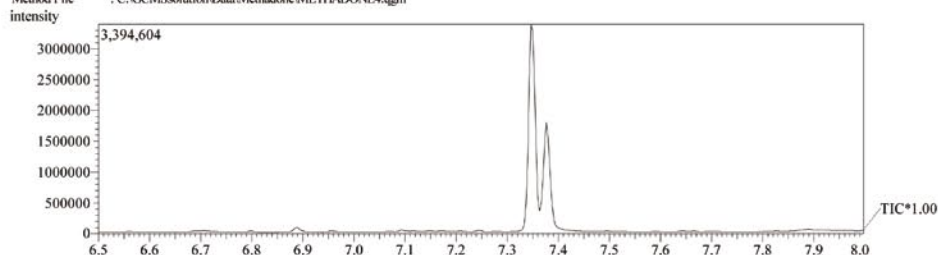
Ions monitored: 303.1, 78.15 for deuterated methadone (d9)
294.0, 72.1 for methadone

Correlation coefficient: Methadone $r^2 = 0.9999446$

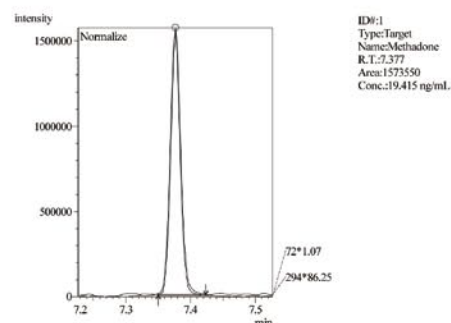
Linearity: 0 – 80 ng/mL; limit of quantitation 10 ng/mL

Data Acquired by : Admin
 Acquisition Date : 7/19/2005 4:07:58 PM
 Sample Name : 20ng/ml
 Data File : D:\Methadone\MED082.qsd
 Method File : C:\GCMSolution\Data\Methadone\METHADONE4.qgm

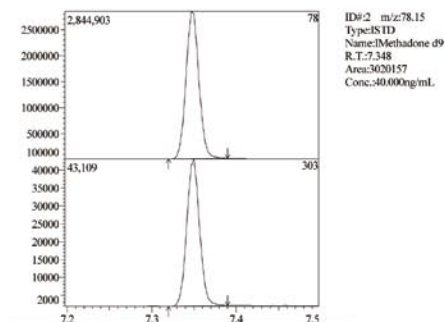
Multicomponent Quantitation Report



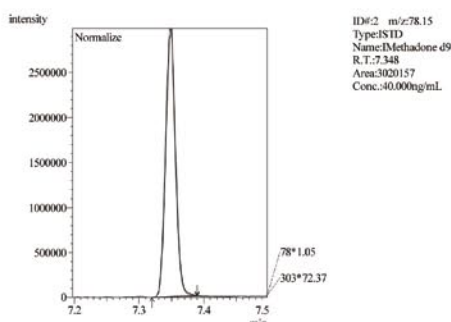
#	m/z	Area	Ratio	Range	Criteria
1	294.00	19499	1.24	0.7 - 1.3	Pass



#	m/z	Area	Ratio	Range	Criteria
1	294.00	19499	1.24	0.7 - 1.3	Pass



#	m/z	Area	Ratio	Range	Criteria
1	303.10	43406	1.44	1.4 - 2.6	Pass



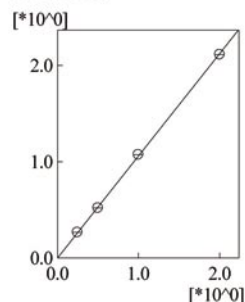
#	m/z	Area	Ratio	Range	Criteria
1	303.10	43406	1.44	1.4 - 2.6	Pass

Analyst _____ Date _____

Approved by _____ Date _____

Printed on: 22 Oct 2007 14:50

ID#1 Mass: 72.15 Name: Methadone
 $f(x) = 1.056983 \cdot x + 0.000000$
 $r^2 = 0.999854$



#	onc.	Ratio (ng/mL)	Mean Area Ratio
1	0.250	0.27	
2	0.500	0.52	
3	1.000	1.07	
4	2.000	2.11	

Calibration
 ID#2 Mass: 78.15 Name: IMethadone d9
 $f(x) = ?$
 $r^2 = 0.000000$

ISTD

About the GCMS-QP2010 Series

- **GCMS-QPQ2010 Plus**
- **GCMS-QP2010S**

The most sensitive GC/MS on the market, Shimadzu's GCMS-QP2010 Plus delivers better performance and reliability with unsurpassed hardware capabilities and powerful, flexible software. Utilizing many of the high-end features of the GCMS-QP2010 Plus, the GCMS-QP2010S offers high throughput and excellent productivity, providing users with an excellent performance-to-cost ratio. The GCMS-QP2010 Series features:

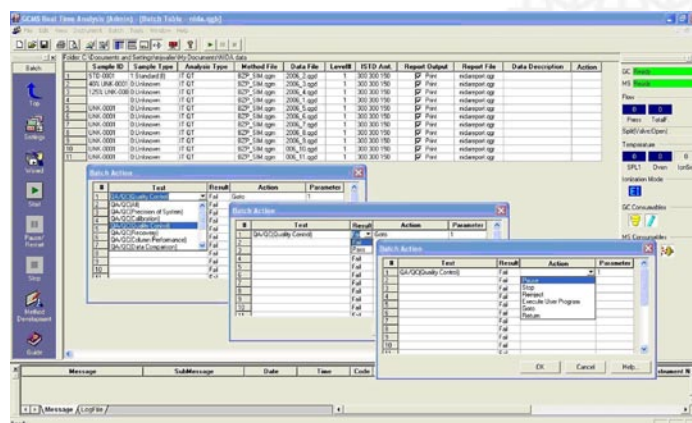
- Extended mass range
- Powerful vacuum
- 20 temperature ramps
- High-performance split-flow TMP
- Patented constant linear velocity

GCMSsolution software combines ease of use and versatility for increased productivity. Features include:

- FASST (Fast Automated Scan/SIM Technology)
- AART (Automatic Adjustment of Retention Time)
- COAST (Creation of Automatic SIM (Scan/SIM) Table)
- Intelligent batch sequencing
- Powerful quantitation
- Flexible, customized reporting

Toxicology Reporting Module

This newly-developed reporting module enables high-throughput drug testing laboratories to address specific productivity and QA/QC requirements. Corrective action plans established by most regulatory methods can be automated and samples may be evaluated during batch sequence injections for a variety of QA/QC parameters. In addition, laboratory productivity is enhanced by automating the data processing and reporting tasks associated with the daily workflow of drug screening analysis.



Regional Offices

MID-ATLANTIC Region
Washington, DC Office
7102 Riverwood Dr.
Columbia, MD 21046
Phone: (410) 381-6996
Toll Free: (800) 388-6996
FAX: (410) 290-9140

NORTH ATLANTIC Region
Boston Office
136 Longwater Drive
Norwell, MA 02061
Phone: (781) 878-7755
Toll Free: (800) 396-4943
FAX: (781) 878-7212

NEW JERSEY Region
New Jersey Office
262 D Old New Brunswick Rd.
Piscataway, NJ 08854
Phone: (732) 981-4400
Toll Free: (800) 439-8555
FAX: (732) 981-4420

SOUTHEAST Region
Raleigh/Durham Office
4022 Stirrup Creek Drive, Suite 312
Durham, NC 27703
Phone: (919) 425-1010
Toll Free: (800) 951-9167
FAX: (919) 544-3497

NORTH CENTRAL Region
Chicago Office
2055 W. Army Trail Rd. Suite 106
Addison, IL 60101
Phone: (630) 916-6286
Toll Free: (800) 792-1992
FAX: (630) 916-7160

SOUTH CENTRAL Region
Houston Office
10801 Hammerly Blvd., Suite 148
Houston, TX 77043
Phone: (713) 467-1151
Toll Free: (800) 739-1942
FAX: (713) 467-1153

MID-WEST Region
Kansas City Office
8052 Reeder
Lenexa, KS 66214
Phone: (913) 888-9449
Toll Free: (877) 698-7923
FAX: (913) 888-8388

NORTHERN CALIFORNIA Region
San Francisco Office
7060 Koll Center Pkwy. Suite 328
Pleasanton, CA 94566
Phone: (925) 417-2090
Toll Free: (800) 482-0253
FAX: (925) 462-7348

SOUTHERN CALIFORNIA Region
Carlsbad Office
1817 Aston Avenue, Suite 105
Carlsbad, CA 92008
Phone: (760) 710-2400
Toll Free: (866) 862-1677
FAX: (760) 931-9854

Founded in 1875, Shimadzu Corporation, a leader in the development of advanced technologies, has a distinguished history of innovation built on the foundation of contributing to society through science and technology. We maintain a global network of sales, service, technical support and applications centers on six continents, and have established long-term relationships with a host of highly trained distributors located in over 100 countries. For information about Shimadzu, and to contact your local office, please visit our Web site at www.shimadzu.com.



SHIMADZU CORPORATION International Marketing Division

3. Kanda-Nishikicho 1-chome, Chiyoda-ku, Tokyo 101-8448, Japan
Phone: 81(3)3219-5641, Fax: 81(3)3219-5710
URL: www.shimadzu.com

SHIMADZU SCIENTIFIC INSTRUMENTS, INC.

7102 Riverwood Drive, Columbia, Maryland 21046, U.S.A.
Phone: 800-477-1227/410-381-1227, Fax: 410-381-1222
URL: www.ssi.shimadzu.com